

# Plant DNA detection from grasshoppers' gut contents: method and applications

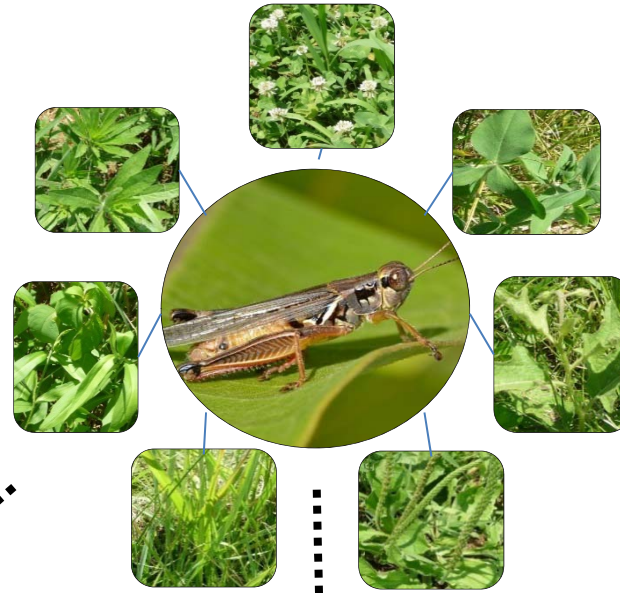
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# Detecting plant meals in insects guts

- understanding insect feeding preferences



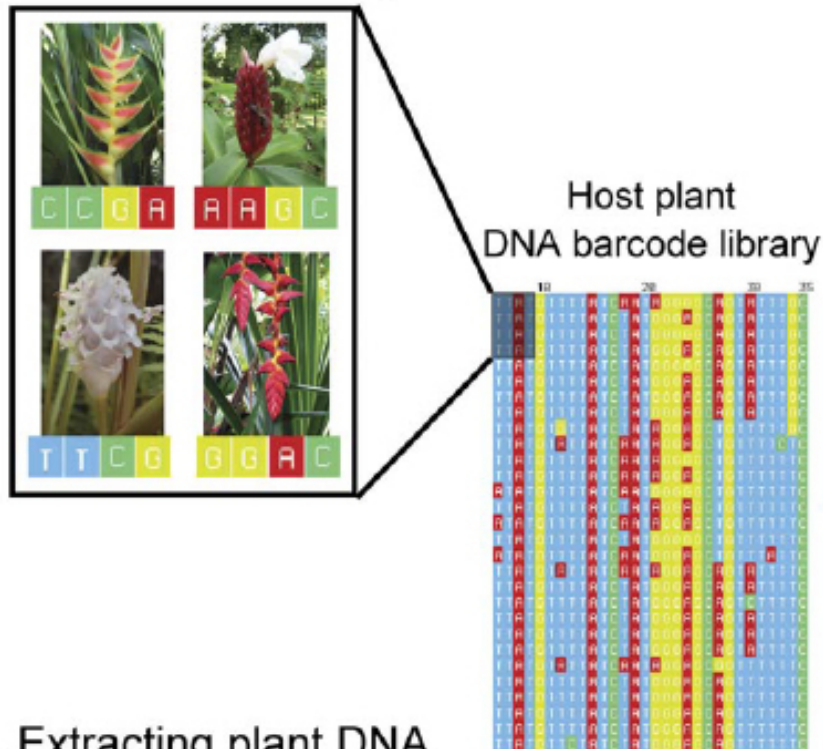
- detecting and predicting plant-insect associations

Accurate confirmation of plant food digestion is critical



# Molecular confirmation of diet

A. Assembling a host plant DNA barcode library

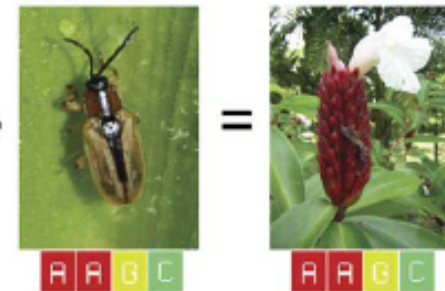


B. Extracting plant DNA from insect herbivores



C. Comparing extracted DNA with sequences in the DNA barcode library

D. Matching DNA sequences and host plant identification



# Existing protocols for plant DNA detection within insect guts

## Limitations:

- Protocols have been developed for a limited number of insect species – **protocols for grasshoppers?**
- Information about insect tissue preparation is limited – **different size of insects?**
- Plant DNA detectability **in different parts of insect digestive system** has not yet been described.



# Research questions

1. Creating the protocol: Which region of plant DNA can be reliably detected in grasshoppers guts?



2. How long can plant DNA be detected in nymphs and adult grasshoppers of different sizes?

3. Is it possible to “follow” the plant DNA during the process of digestion?

# Study species

Nymphs

*Melanoplus sp.*



Adults

*Melanoplus femurrubrum*



*Melanoplus differentialis*



*Miscanthus sinensis*



*Plantago lanceolata*



*Bothriochloa bladhii*



*Trifolium repens*



*Cichorium intybus*



*Bouteloua curtipendula*

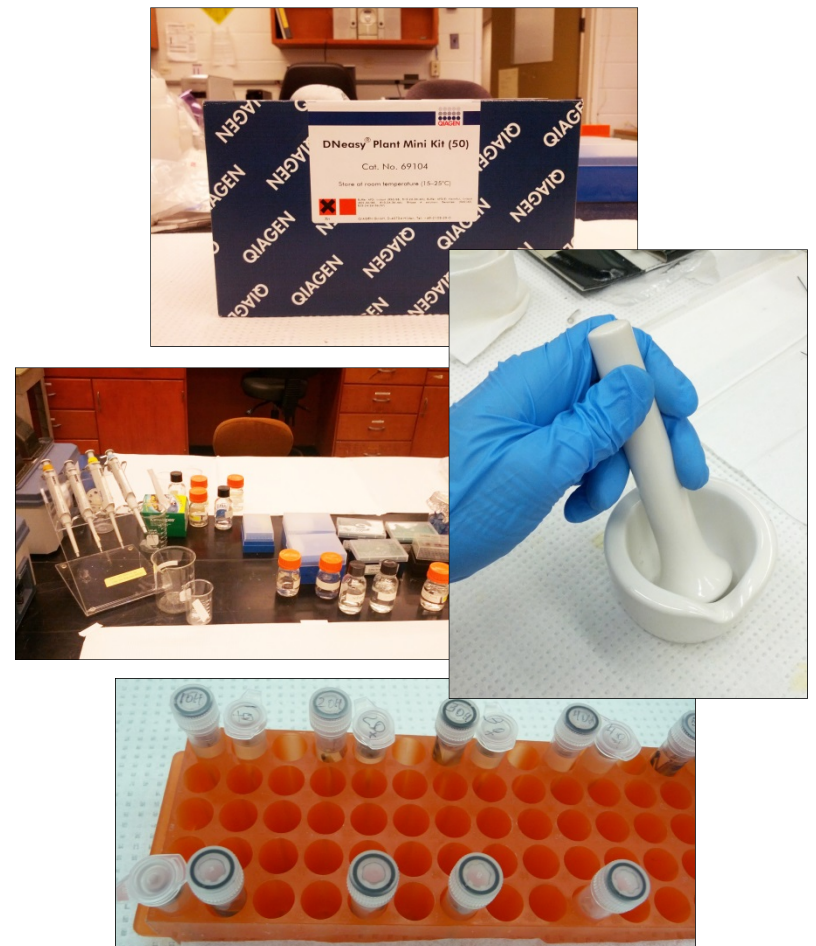


# Creating a protocol for plant DNA detection within grasshopper guts

## Step 1. Dissecting grasshoppers and isolating guts



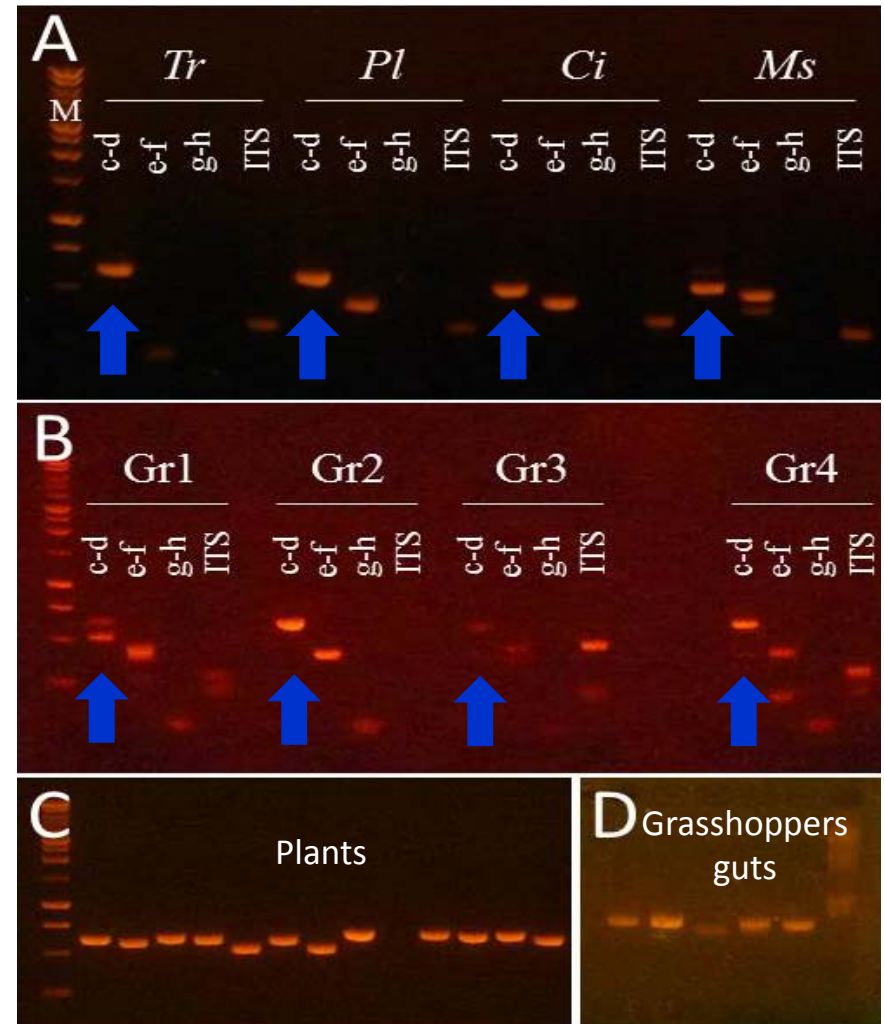
## Step 2. Plant DNA extraction from guts



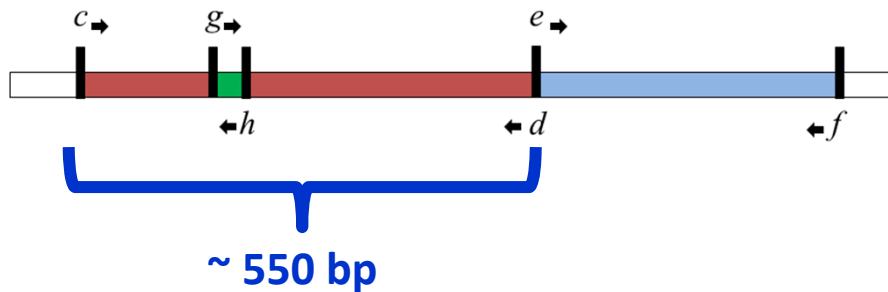
## Step 3. PCR amplification: testing primers

Name	Target region	Sequence 5'-3'	bp
c	trnL (UAA) intron	CGAAATCGGTAGACGCTACG	500
d	trnL (UAA) intron	GGGGATAGAGGGGACTTGAAC	500
e	trnL (UAA)3'exon/trnF(GAA)	GGTTCAAGTCCTCTATCCC	400
f	trnL (UAA)3'exon/trnF(GAA)	ATTTGAACTGGTGACACGAG	400
g	P6 loop of trnL (UAA)	GGGCAATCCTGAGCCAA	40
h	P6 loop of trnL (UAA)	CCATTGAGTCTCTGCACCTAT	40
ITS1F	Nuclear, ITS	C TCCGTAGGTGAACCTGCGG	290
ITS2R	Nuclear, ITS	GCTGCGTTCTTCATCGATGC	290

Plants (A)



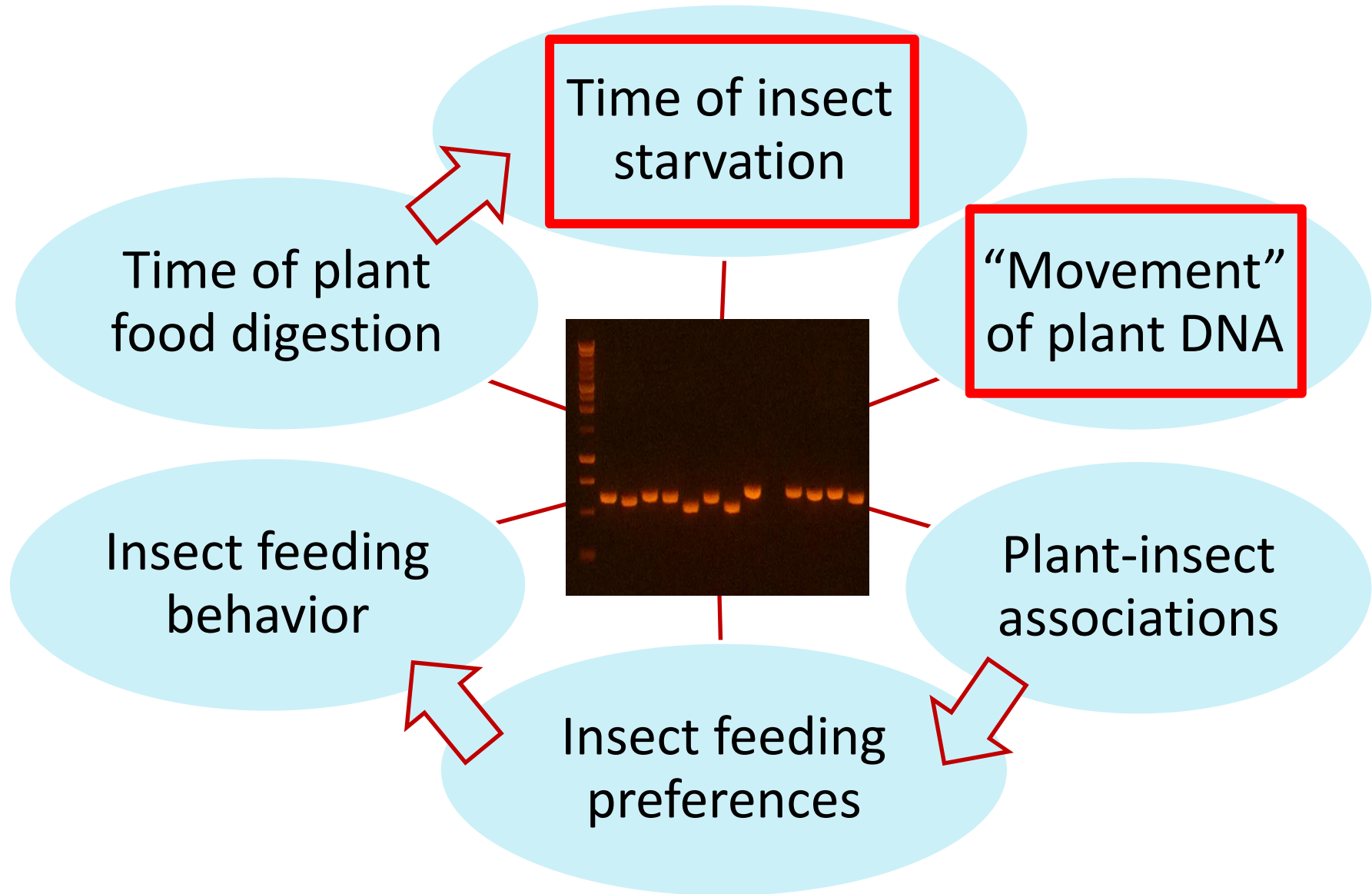
Grasshoppers guts (B)






Primers c-d (C, D)



# Applications of the protocol



# Time of starvation: How long can plant DNA be detected in nymphs and adult grasshoppers of different sizes?

Grasshopper species	Life stage	Weight (g), Mean±1SE	Type of feeding experiment	Plant species used for feeding	Total time of feeding	Tissues for DNA extraction	Time intervals post ingestion (hours)	Plant DNA detectability (hours PI)
<b><i>Melanoplus spp.</i></b> 	Nymph	0.11 ± 0.02	Choice	<i>Bouteloua curtipendula</i> <i>Bothrichloa bladhii</i>	3.5 h	Whole body	0, 2, 4, 6, 8, 10, 12	12 h
<b><i>Melanoplus femurrubrum</i></b> 	Adult	0.35 ± 0.02	No-choice	<i>Bothrichloa bladhii</i>	3.5 h	Whole gut	0, 2, 4, 6, 8, 10, 12	12 h
		0.36 ± 0.01	Choice	Plant mixture	2 d		0, 2, 4, 6, 8, 10	
<b><i>Melanoplus differentialis</i></b> 	Adult	1.66 ± 0.27	Choice	<i>Bouteloua curtipendula</i> <i>Bothrichloa bladhii</i>	3.5 h	Foregut and [midgut+ hindgut] separately	0, 1, 3, 22	22 h

# Plant DNA detectability



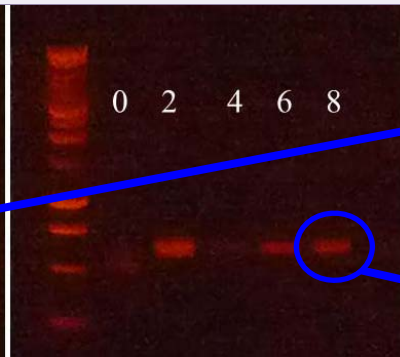
Nymphs *Melanoplus* spp.

12 h PI: choice, two plants

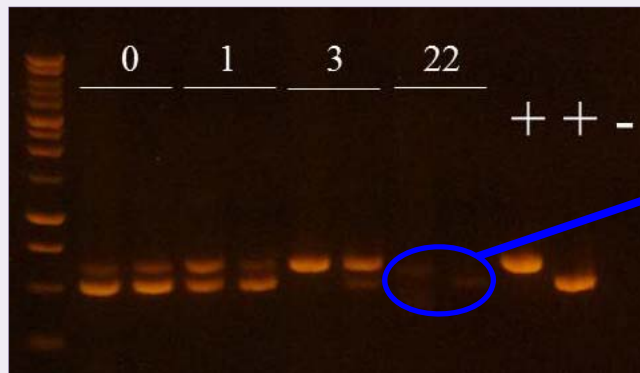


*Melanoplus femurrubrum*

12 h PI: no choice, single plant



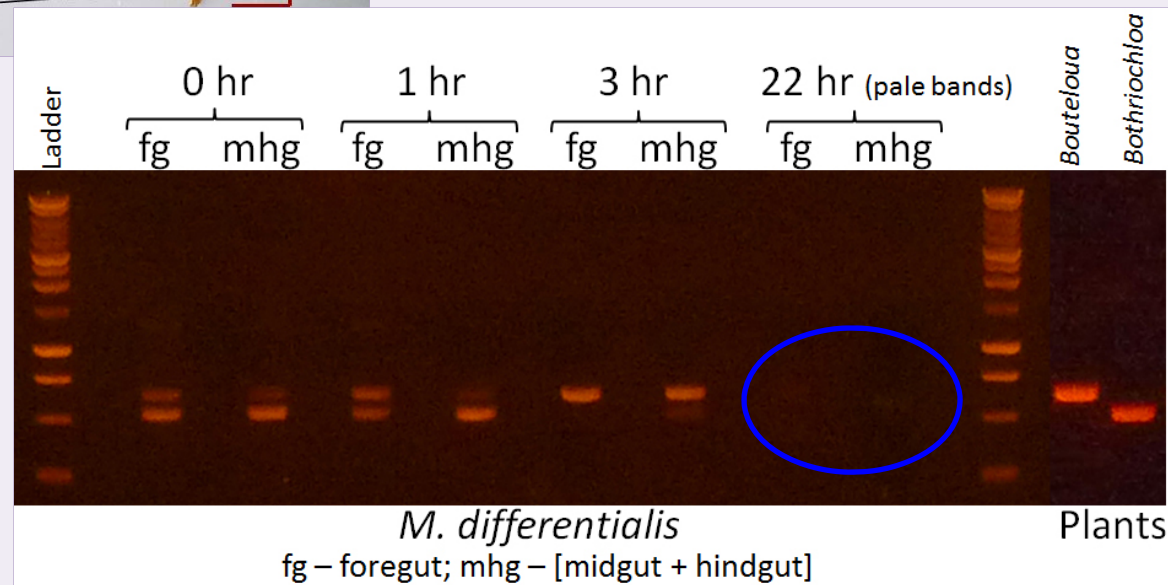
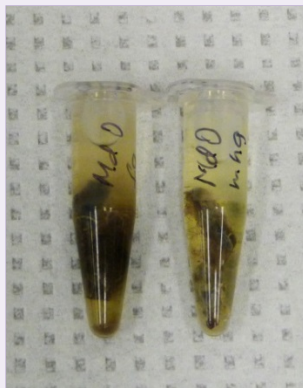
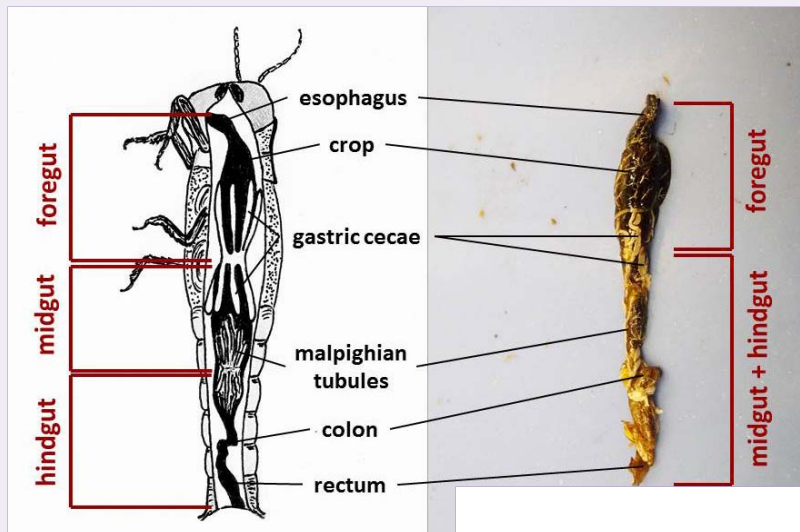
8 h PI: choice, plant mixture



*Melanoplus differentialis*

22 h PI: choice, two plants

# Is it possible to “follow” the plant DNA during the process of digestion?



# Conclusions

- We have developed an effective protocol for plant DNA detection from grasshopper guts: fragments (~550 bp) of the non-coding region of the chloroplast *trnL* (UAA) gene were successfully amplified.
- We have demonstrated the utility of this protocol for determining time of insect starvation before feeding experiments and for detecting plant food “movement” in an insect’s digestive system.

# Future directions

Using the developed protocol:

- Determine plant-insect associations with regard to native and exotic plants
- Explore whether the plant coverage (for both native and exotic plants) affects grasshopper feeding choice





***Thank you!***



**University of Cincinnati:**

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