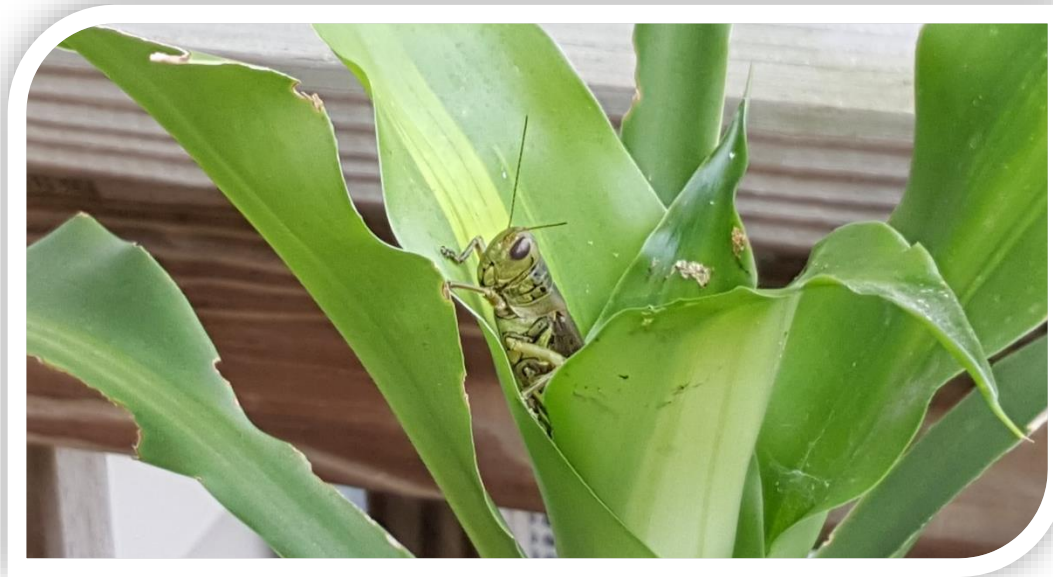


# Invasion Ecology



Week 9

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Narrowing down the focus: novel  
plant-insect interactions

# Learning Outcomes

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***By the end of today's introduction lecture you will know:***

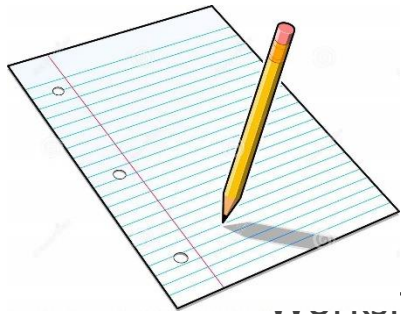
- what novel species associations are
- what mechanisms underlie novel plant-insect interactions

***By the end of today's introduction lecture you will be able to:***

- recognize novel species associations
- give at least two examples of novel plant-insect associations

# Class Activities

---



Worksheet



Small group discussion



All-class-discussion



In-class ungraded quizzes

# Novel Plant-Insect Associations

- a combination of resident (native) and non-resident (exotic) plant or insect species “in which at least one species has **little or no experience with relevant ecological traits of its interaction counterpart**” (Saul and Jeschke, 2015).



Introduced plant's  
native range

Lack of coevolution



Introduced plant



Native insect

Native community

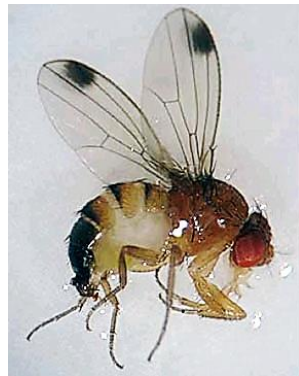
# Diversity of non-native insect and plant species: forest and crop pests

---

➤ Gypsy moth



➤ Spotted Wing Drosophila



➤ Emerald Ash Borer

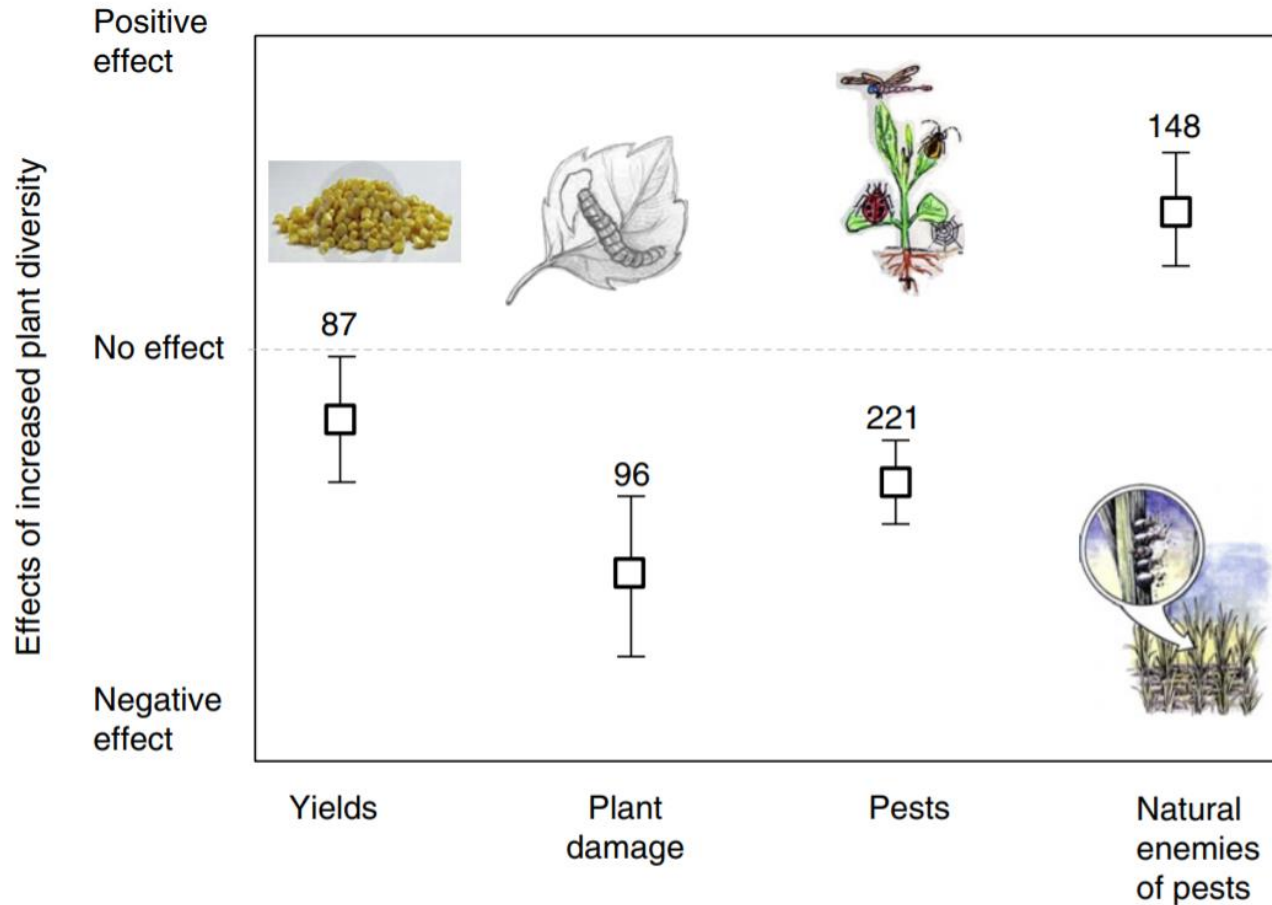


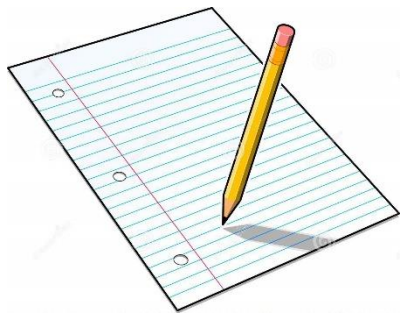
➤ Sirex Woodwasp



# Discussion

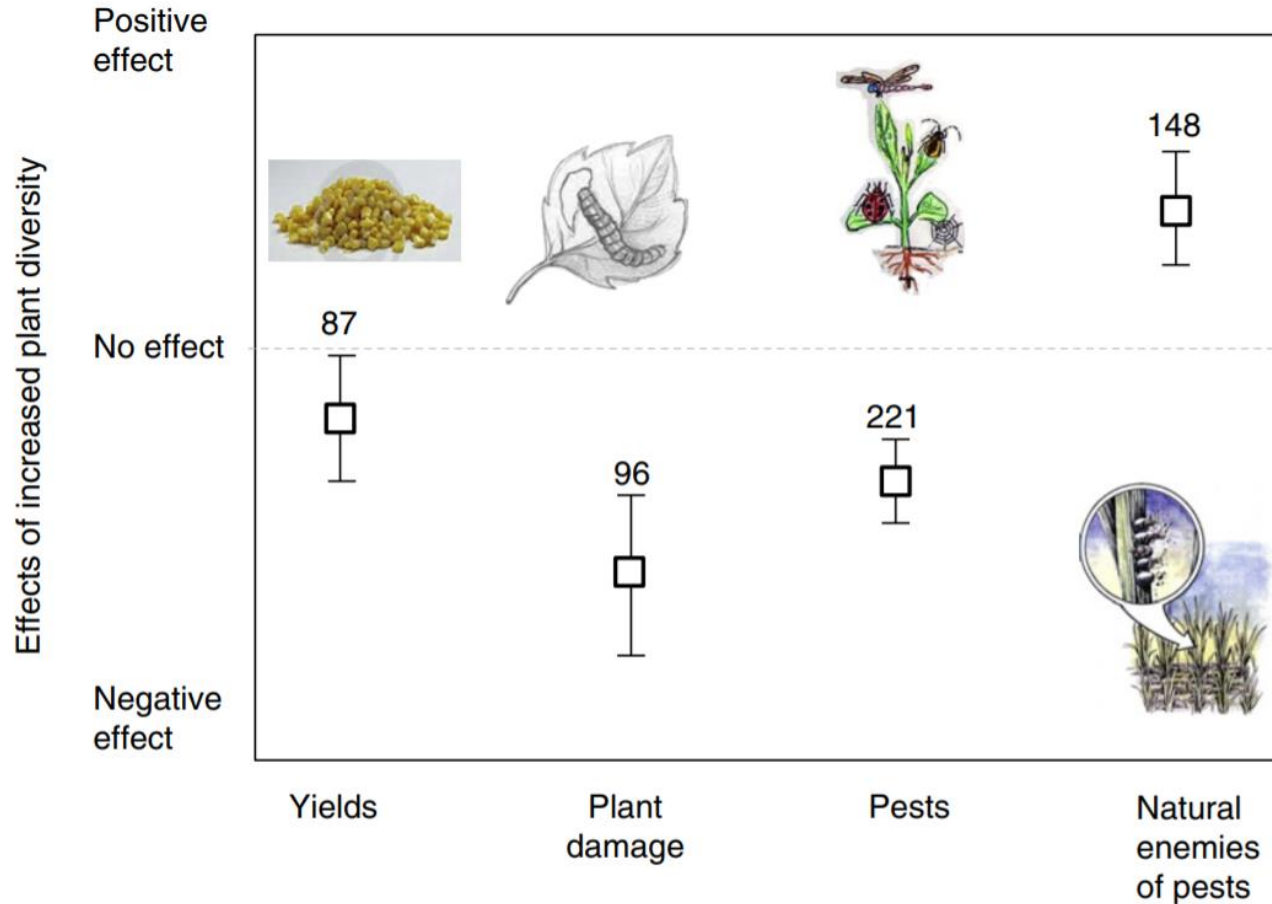
# What do you see?





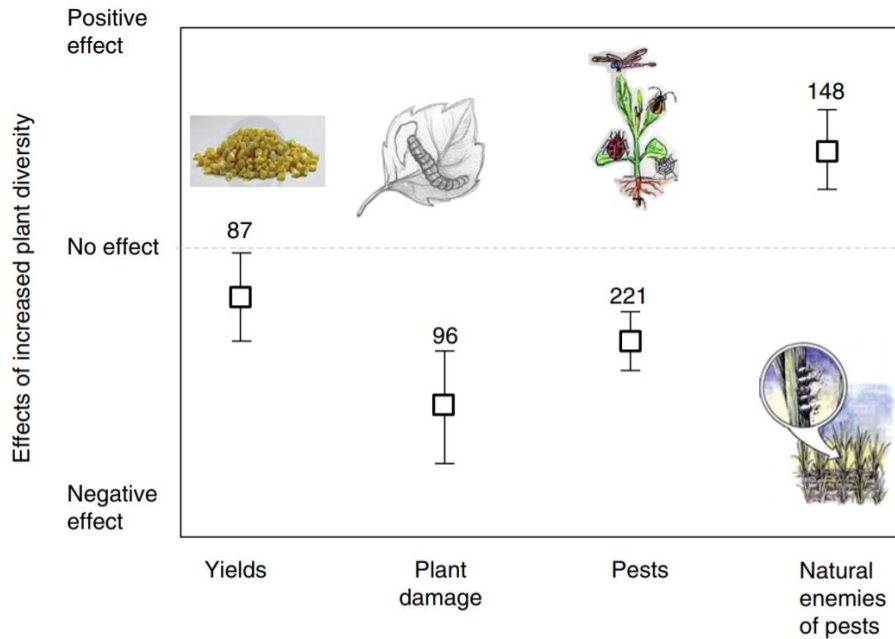
## Worksheet Part 1.

# What does it mean?





# What does it mean?



## Effects of biodiversity:

- lower/high abundance of pests (herbivores) (Yes/No)
- lower/higher abundance of natural enemies of such pests (predators and parasites) (Yes/No)
- Increased/decreased damage by pests (Yes/No)
- Reduction/increase in crop yield (Yes/No)





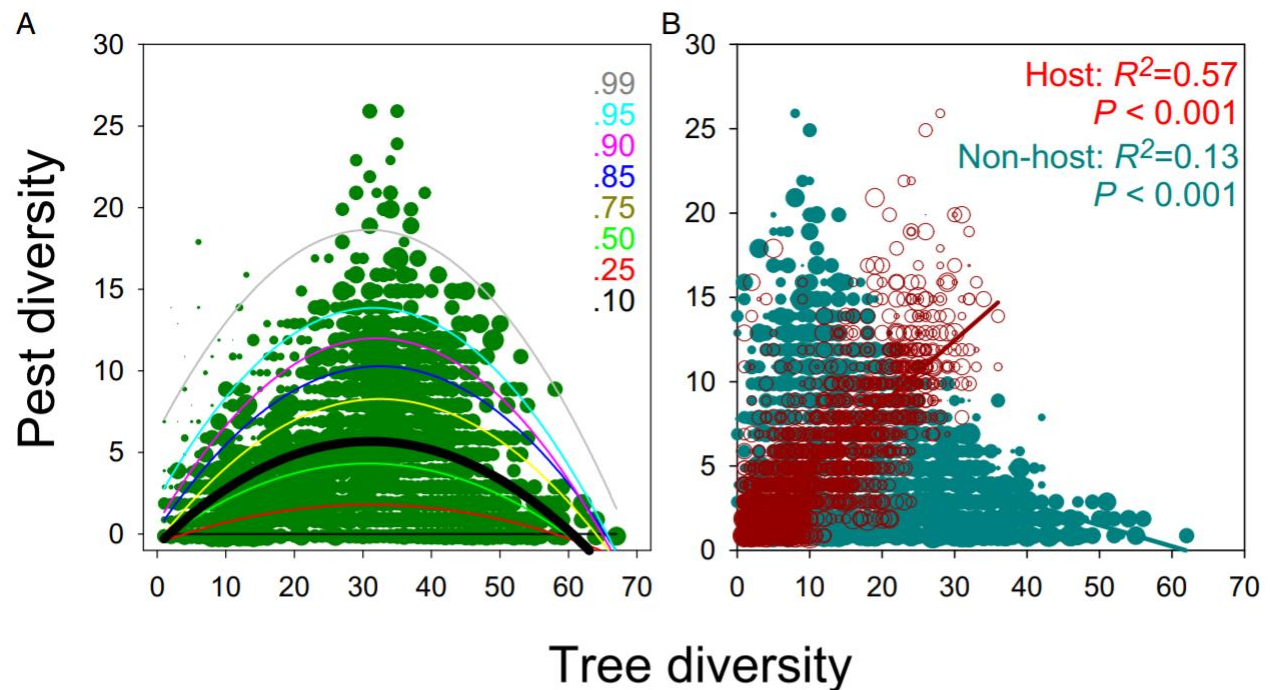
# Tree diversity regulates forest pest invasion

Qinfeng Guo<sup>a,1</sup>, Songlin Fei<sup>b,1</sup>, Kevin M. Potter<sup>c</sup>, Andrew M. Liebhold<sup>d,e</sup>, and Jun Wen<sup>f</sup>

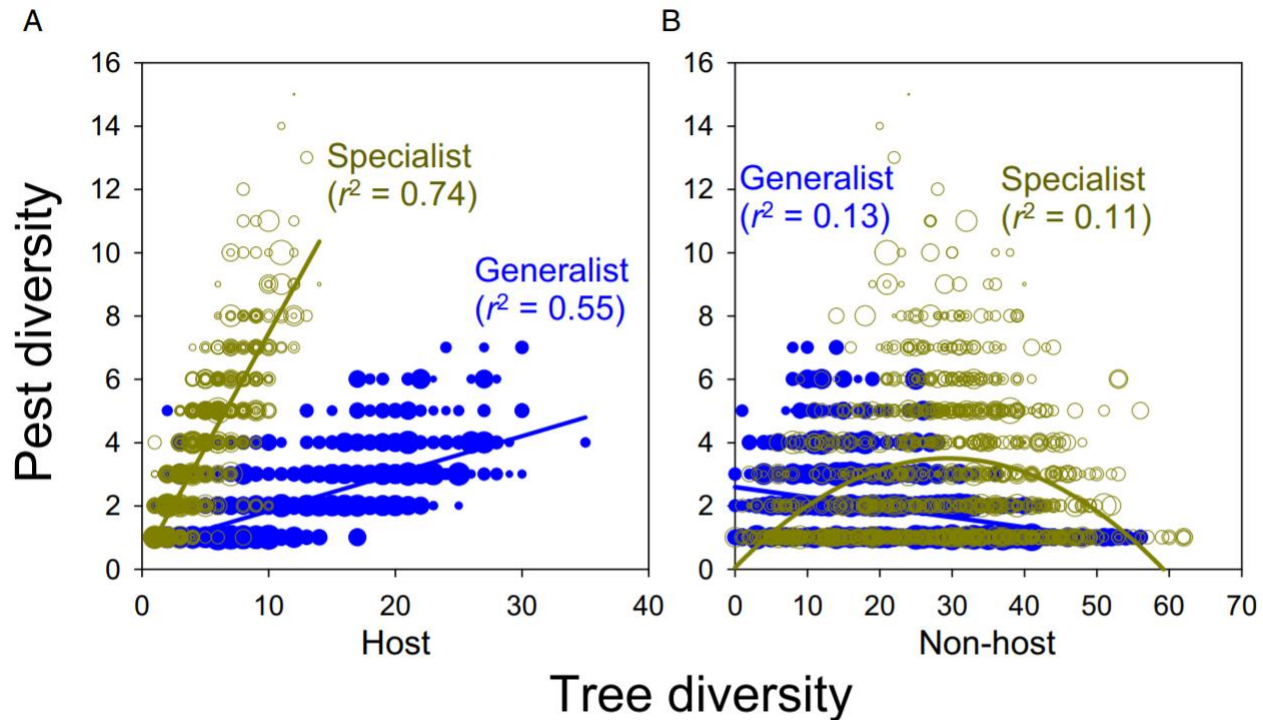
<sup>a</sup>US Department of Agriculture Forest Service, Southern Research Station, Eastern Forest Environmental Threat Assessment Center, Research Triangle Park, NC 27709; <sup>b</sup>Department of Forestry and Natural Resources, Purdue University, West Lafayette, IN 47907; <sup>c</sup>Department of Forestry and Environmental Resources, North Carolina State University, Research Triangle Park, NC 27709; <sup>d</sup>US Department of Agriculture Forest Service, Northern Research Station, Morgantown, WV 26505; <sup>e</sup>Faculty of Forestry and Wood Sciences, Czech University of Life Sciences Prague, CZ 165 21 Praha 6-Suchdol, Czech Republic; and <sup>f</sup>Duke Clinical Research Institute, Duke University, Durham, NC 27705

Edited by Rodolfo Dirzo, Department of Biology, Stanford University, Stanford, CA, and approved March 5, 2019 (received for review December 10, 2018)

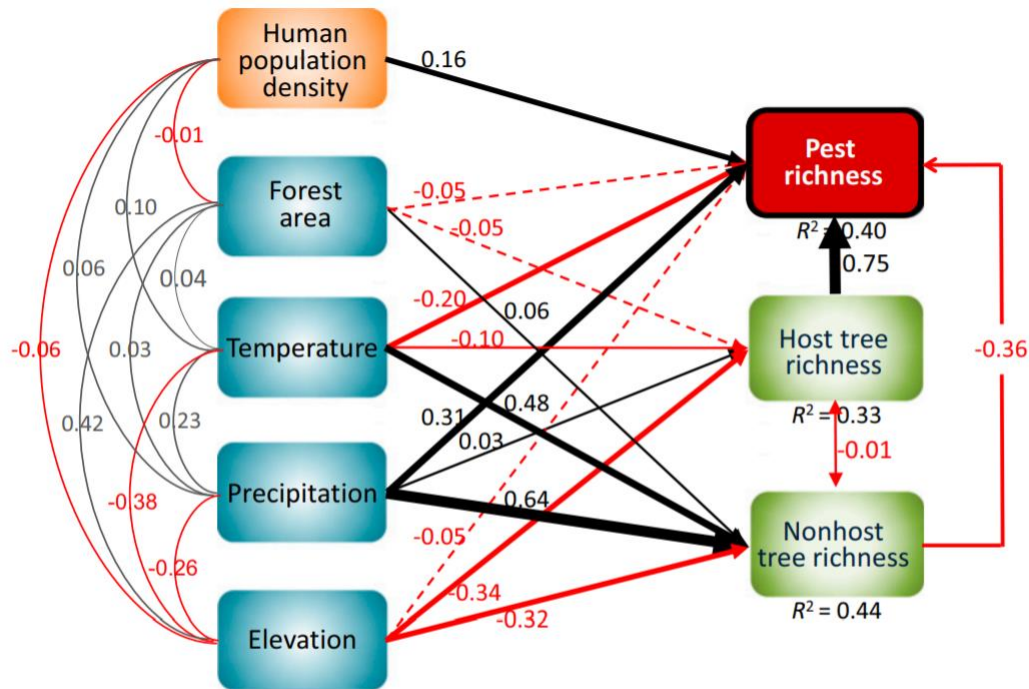
**Nonnative pests often cause cascading ecological impacts, leading to detrimental socioeconomic consequences; however, how plant** are (e.g., host vs. nonhosts and their relative proportions), as well as on the direct and indirect interactions among neighboring



# Native host/nonhost diversity and the diversity of specialist and generalist nonnative invasive pests in forest ecosystems



# The effects of selected physical and human factors on pest diversity





# Quick Review (true/false)

---

- Novel plant-insect associations are the ones between invasive insects and native plants only **Yes/No**
- Plant diversity increases the number of natural enemies **Yes/No**
- Tree diversity affect generalist insects only **Yes/No**
- Temperature doesn't affect invasive pest diversity **Yes/No**

# Interactions between native and invasive species and their ecological impact: a case study

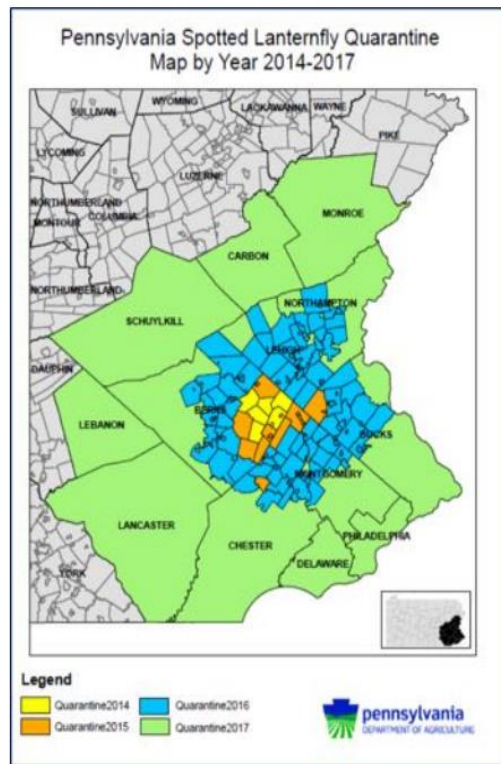
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- **The spotted lanternfly**
- An emerging highly invasive insect pest
- Native to China
- Invaded Korea in 2004
- Detected in Berks County, PA in 2014

# Where is spotted lanternfly in the US and how fast will it spread?

## Invasion process



Pennsylvania – Berks (2014) now in 13 counties, established

Delaware – New Castle (2017), established

New Jersey – Hunterdon, Mercer and Warren Counties (2018) established

New York – Albany, Suffolk and Yates (2018)

Virginia - Frederick County (2018), established

Massachusetts (2019)

# Life stages of spotted lanternfly

---



Eggs

1<sup>st</sup>  
instar

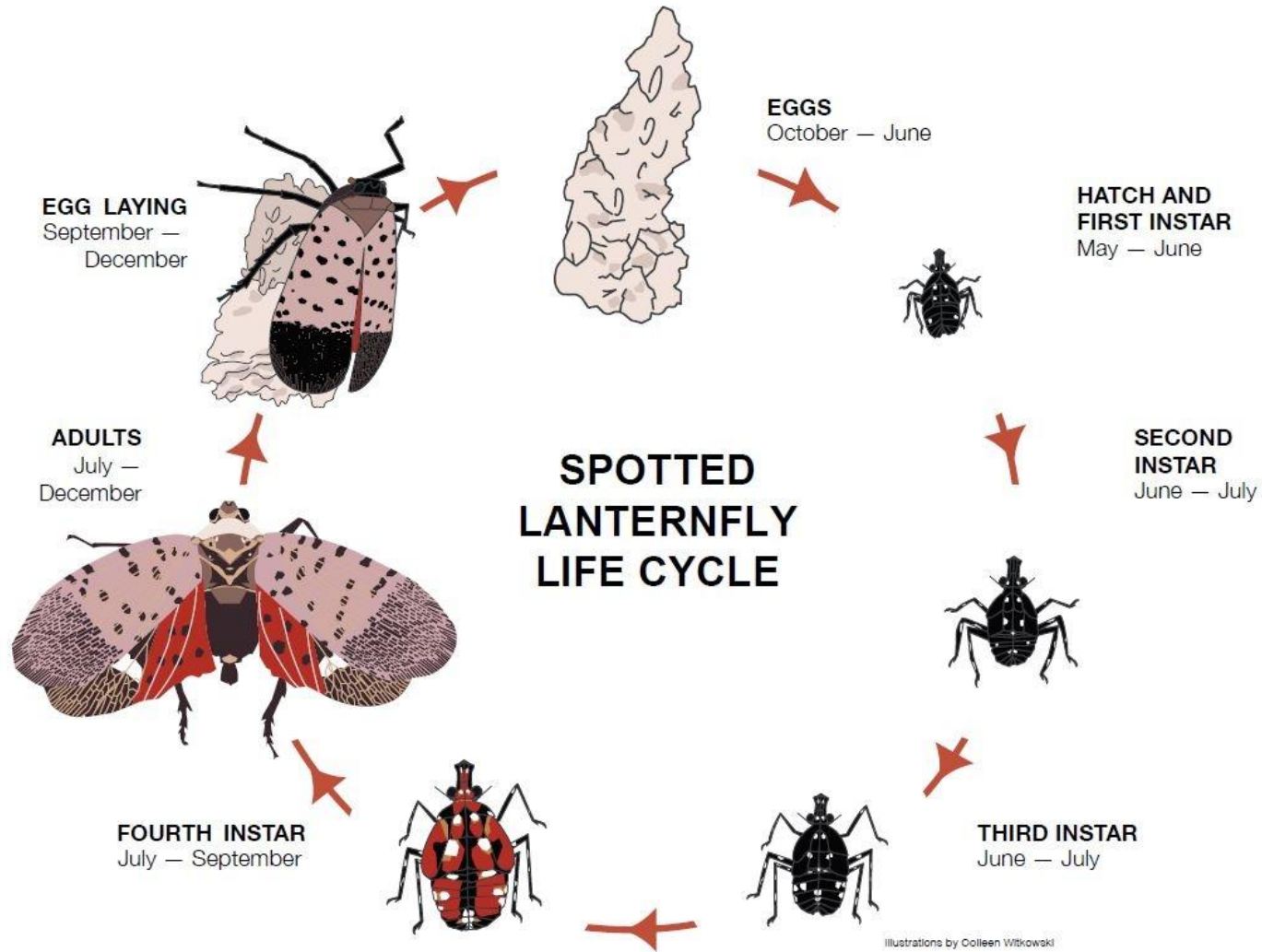
2<sup>nd</sup>  
instar

3<sup>rd</sup>  
instar

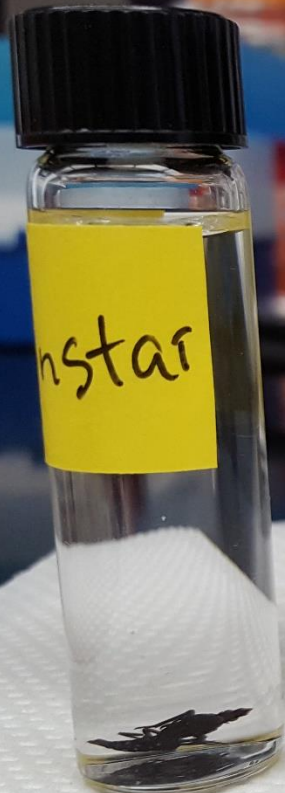
4<sup>th</sup>  
instar

Adult

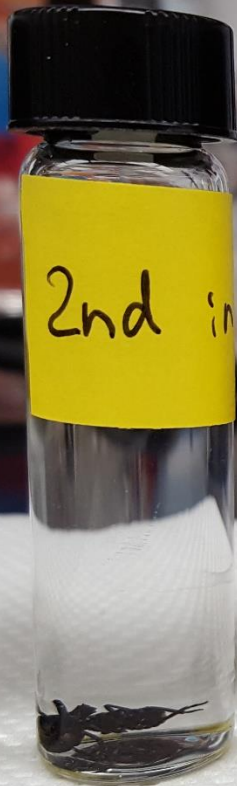
# Life cycle of spotted lanternfly



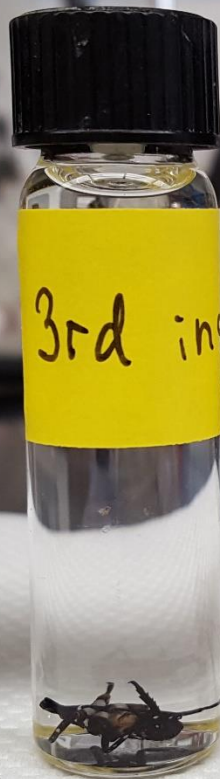




1<sup>st</sup>  
instar



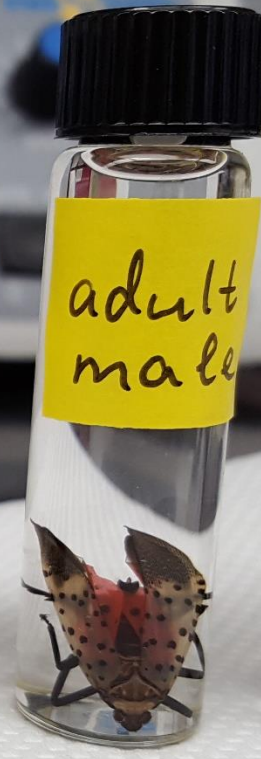
2<sup>nd</sup>  
instar



3<sup>rd</sup>  
instar



4<sup>th</sup>  
instar



Adult  
male



Adult  
female

# Modes of SLF dispersal

---

- One of the most aggressive leaf-hopping pest in Mid-Atlantic region
- Very high potential to breed and increase its population size
- It can be spread long distances by any material (including manmade material) containing egg masses:

- ❖ trunked tree
- ❖ stones
- ❖ vehicles
- ❖ yard furniture
- ❖ farm equipment, etc.

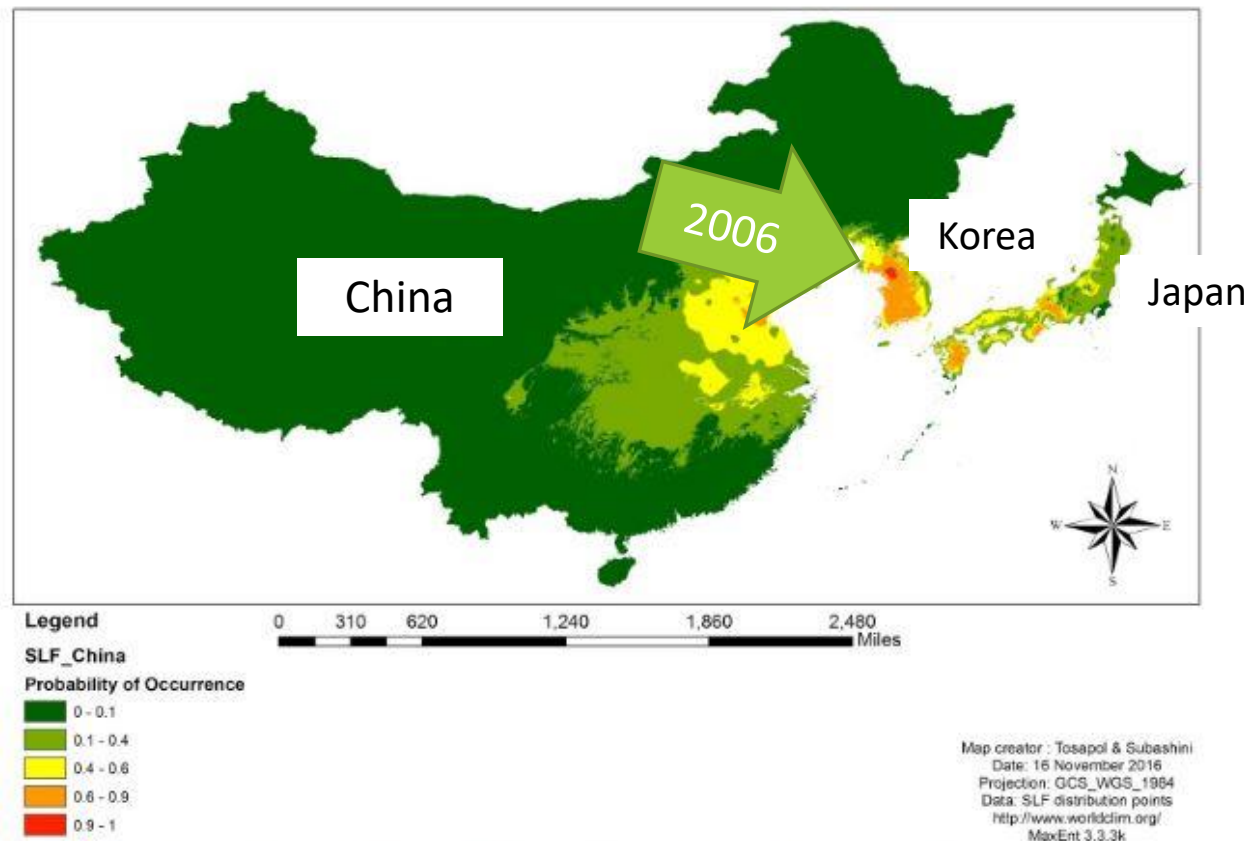


# What are the native and invaded ranges of spotted lanternfly in Asia?

Native to  
China

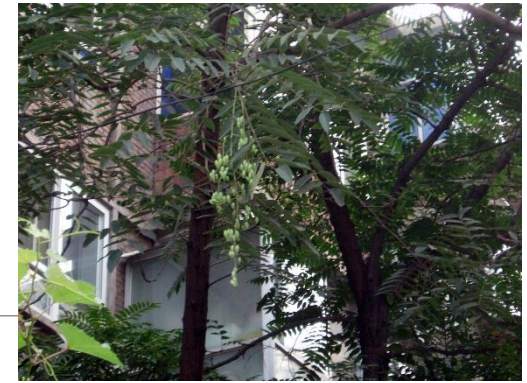
Invasive in  
Korea, Japan,  
Taiwan, and  
Vietnam

The predicted probability distributions of Spotted Lanternfly in China, Korea and Japan



Map: The predicted probability distributions of Spotted Lanternfly in China, Korea and Japan

# Host plants in China



Chinese mahogany



Tree-of-heaven



Manchurian catalpa



'Chinese Elm'

- Ornamental and fruit trees
- Soybean and some agricultural crops



Soybean plants

# Host plants in Korea

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Tree-of-heaven



Manchurian walnut



Amur grape



Chinese mahogany



Korean Evodia

- Ornamental and fruit trees
- Herbs

# Egg masses



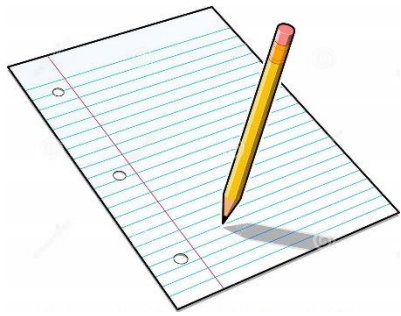
# Host plants

Sap-feeder

SLF can utilize over 70 host plants:

- **Apple**
- Plum
- Cherry
- Peach
- Apricot
- **Grape**
- Pine
- Tree of heaven (preferred tree host)
- ..... and many many others....





## Worksheet Part 2.

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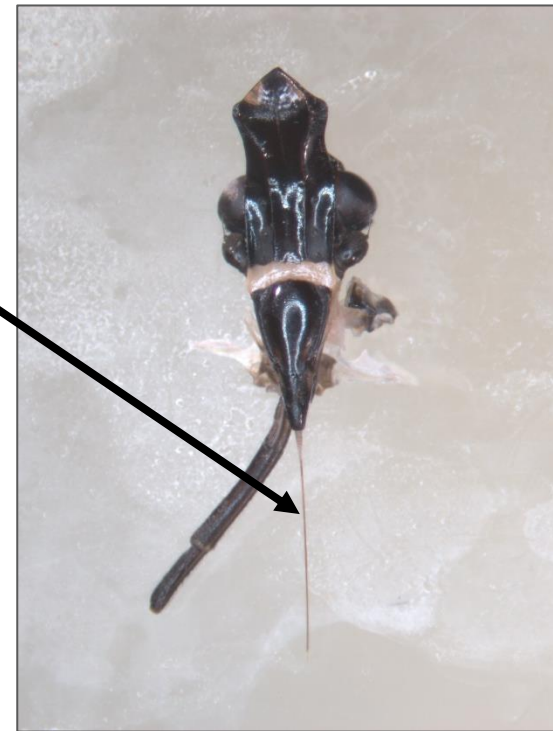
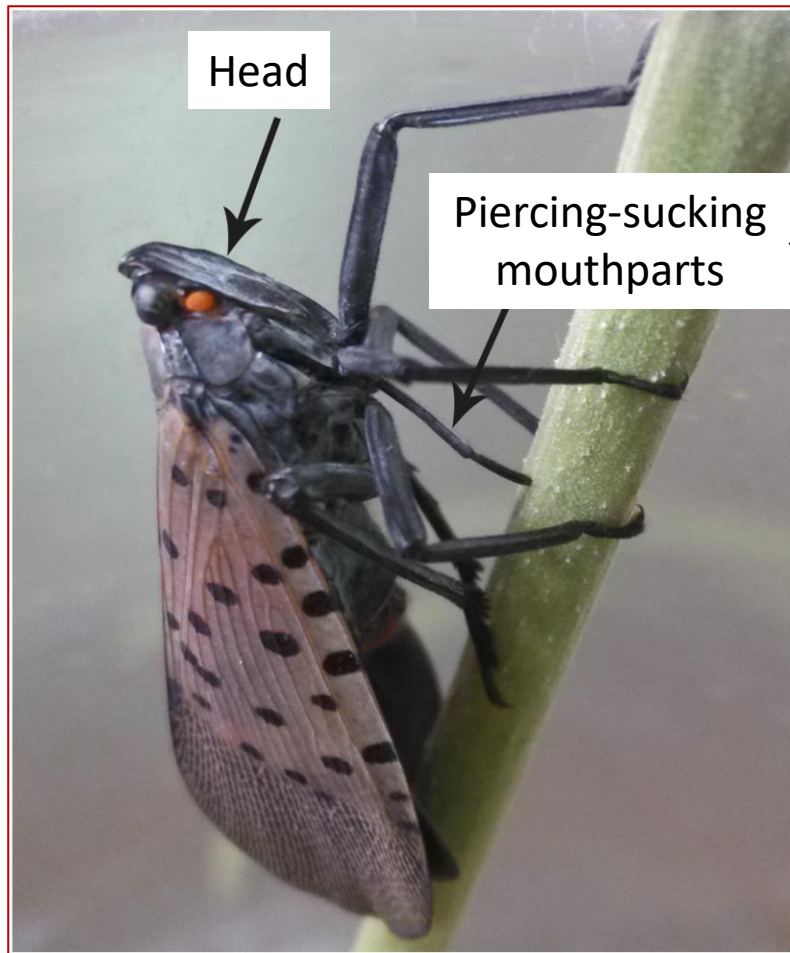
Please list 2 native and 2 novel host plants for the spotted lanternfly  
(1-2 min)



Please compare your lists in pairs and create a combined list  
(1-2 min)



# How does the lanternfly eat and damage plants?



Sap- feeders

# Plant damage

Consumes  
phloem sap

Reduction in  
photosynthesis

Weeping wounds

Decreasing  
plant's growth



# Plant damage



Create a sugary substance (honeydew)

Attract other insects - ants, wasps, etc.

Colonized by sooty mold -> blackening of parts of the plant











# Behavior on host trees

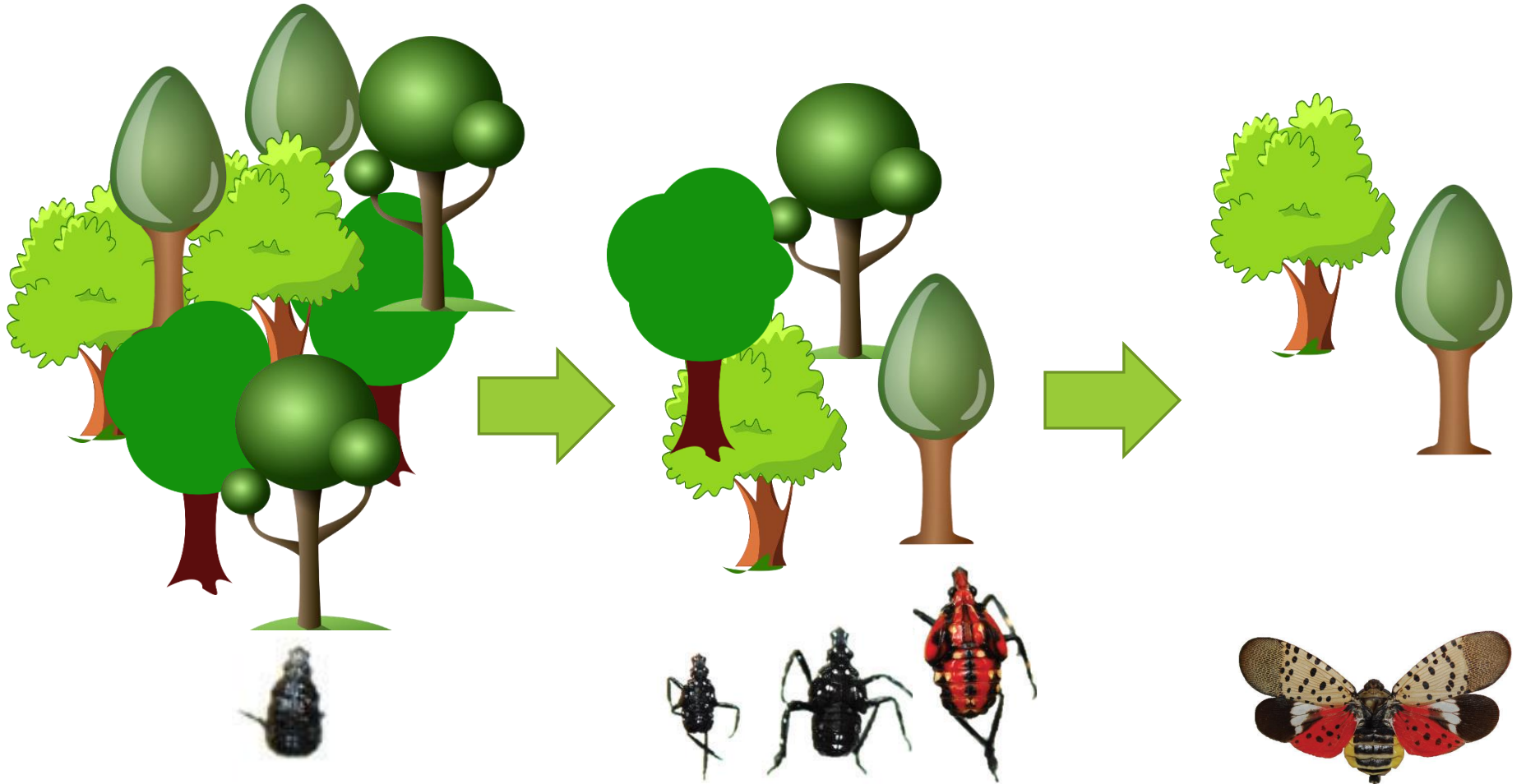
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# Seasonal behavior

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May-June

June-August

September-December

# Behavior on host trees

---

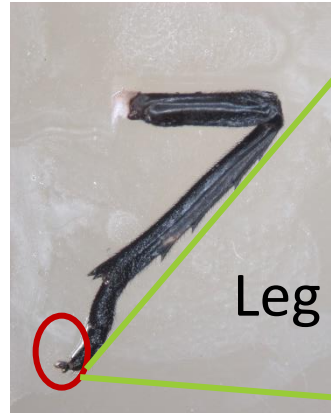


# Behavior on host trees

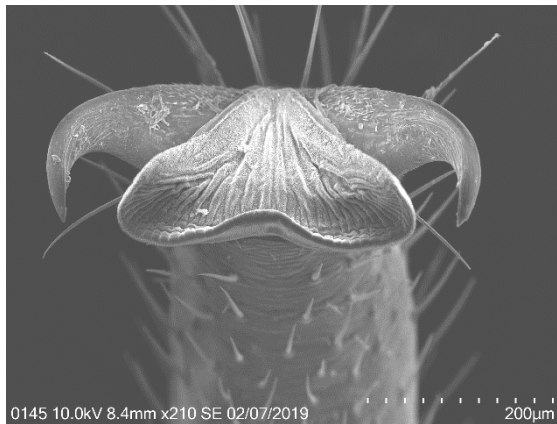
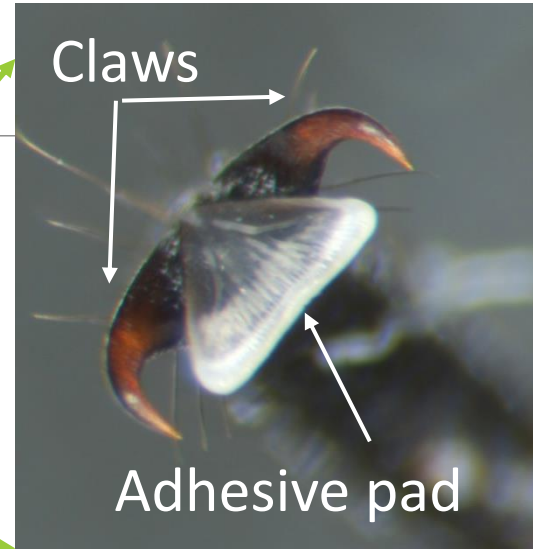
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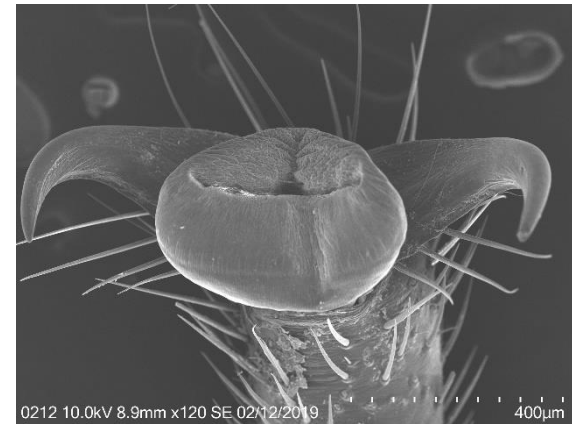
# Holding on to the host trees



Leg tip



Lanternfly resting



Lanternfly moving



# Quick Review

---

- In the introduced range, does the spotted lanternfly feed on host plants present in its native range only? Yes/No
- Adaptations to feeding on novel host plants?
  - seasonal behavior Yes/No
  - morphological adaptations Yes/No
  - nymphal coloration Yes/No

# Management strategies in PA



## Band trees to catch nymphs

### Did you know?

In the spring, spotted lanternfly nymphs crawl up trees to find a place to feed— stop them by banding trees with sticky paper or tape.



Pennsylvania Department of Agriculture



PennState

## Scrape eggs

Eggs should be scraped off of trees, posts, stones, houses, and anywhere else you find them!

Use a plastic card, putty knife, or stick to scrape eggs downward into a bottle or bag.

Eggs can then be killed by putting them in rubbing alcohol, smashing them, or burning them.

Removes 30-50 eggs per mass

Link to video:

<https://extension.psu.edu/how-to-remove-spotted-lanternfly-eggs>



PA Dept. Ag.



PennState

# Biological control

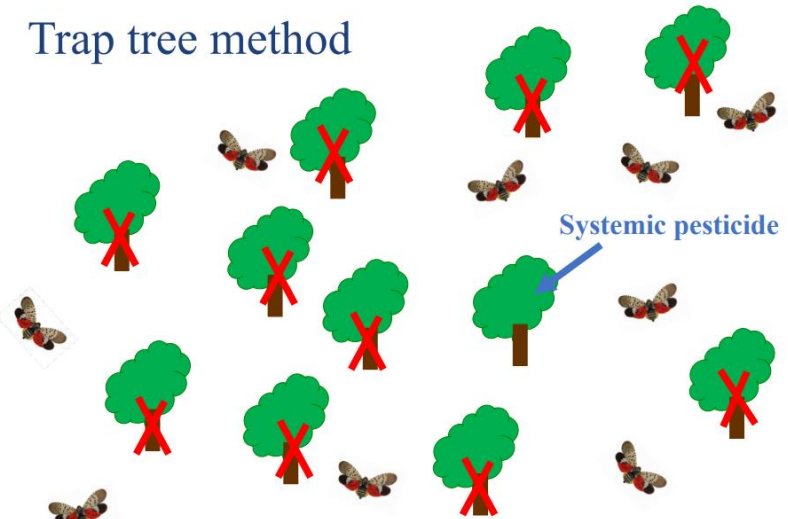
Indigenous natural enemies including spiders, mantises, and assassin bugs are now attacking and killing lanternflies



A tiny wasp called *Ooencyrtus kuvanae* was imported in 1908 to control gypsy moth. It was taken a liking to spotted lanternfly and now parasitizes and kills eggs of the lanternfly.

# Use trap-trees to reduce populations

Trap tree method



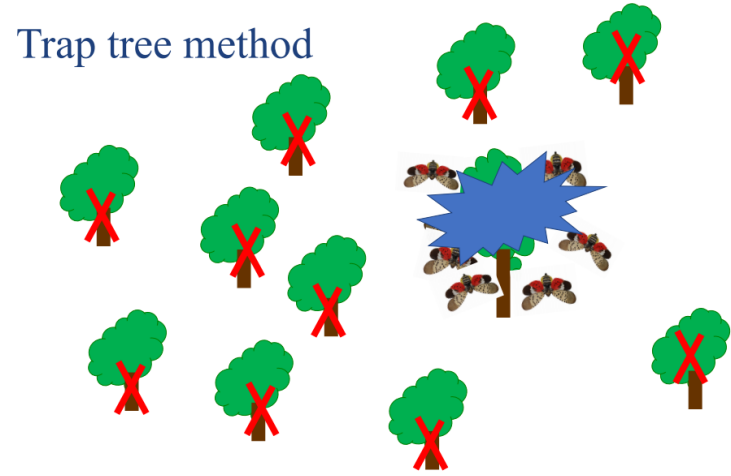
Systemic pesticide

E. Swackhamer & A. Corman

PennState

This diagram illustrates the initial setup of the trap tree method. It shows a field of approximately 15 green trees, each marked with a red 'X' to indicate they are trap trees. A central tree is being treated with a systemic pesticide, indicated by a blue arrow and the text 'Systemic pesticide'. Several butterflies are scattered throughout the field, representing the pest population.

Trap tree method



E. Swackhamer & A. Corman

PennState

This diagram illustrates the result of the trap tree method. The central tree, which was previously treated with systemic pesticide, now has a large blue starburst around it, indicating a high concentration of butterflies. The other trees in the field are still marked with red 'X's, but they are no longer attracting butterflies, showing that the pest population has been effectively reduced to the trap tree.



# Use trap-trees to reduce populations



# Monitoring and scouting



**Egg masses:** on tree trunks, stones, etc.



**Adults:** in clusters on tree trunks



**2-4<sup>th</sup> instars:** plant leaves, stems, tree trunks

**1<sup>st</sup> instar:** close to the ground, plant shoots, stems, etc.

# Insecticidal control

---



Synthetic pyrethroid - deltamethrin 1% EC

Organophosphate - fenitrothion 50% EC

“Quick and strong insecticidal activity against the 2nd-3rd nymphs”

Neonicotinoids - imidacloprid 4% SL and clothianidin 8% SC “showed 100% insecticidal activity at 24h after treatment”

Park et al. 2009

Pyrethrum, Sophora, and neem extracts (at 1,000 fold dilution) killed 95% of adults within 48 h, but the extracts tended to be less effective against nymphs in some tests

Dara et al. 2015

# What can be done to assist with management of lanternfly?

---

Pennsylvania Department of Agriculture and Penn State web sites assist citizens with identification of this new pest, learning how to destroy egg masses, and for reporting sightings in general.

<https://extension.psu.edu/spotted-lanternfly>



If you discover an egg mass, nymphs, or adult lanternflies, report to your University Extension Service or State Department of Agriculture.

<http://extension.umd.edu/hgic/topics/spotted-lanternfly>





## Quick discussion (3 min)

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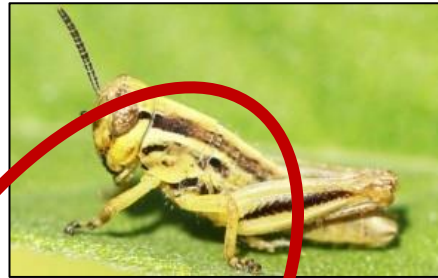


Can you predict how other invasive insects might adapt to their novel host plants?

# Novel plant-associations: implications for co-evolution, biotic resistance, and biological control

**Novel  
association**

Native insect  
herbivore



**Introduced plant**

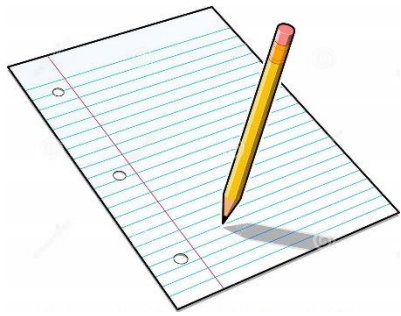


**Native plants**

Native community

# In the introduced range...





## Worksheet Part 3.

---



Why do introduced species fail to establish in a new range?





## Quick discussion (3 min)

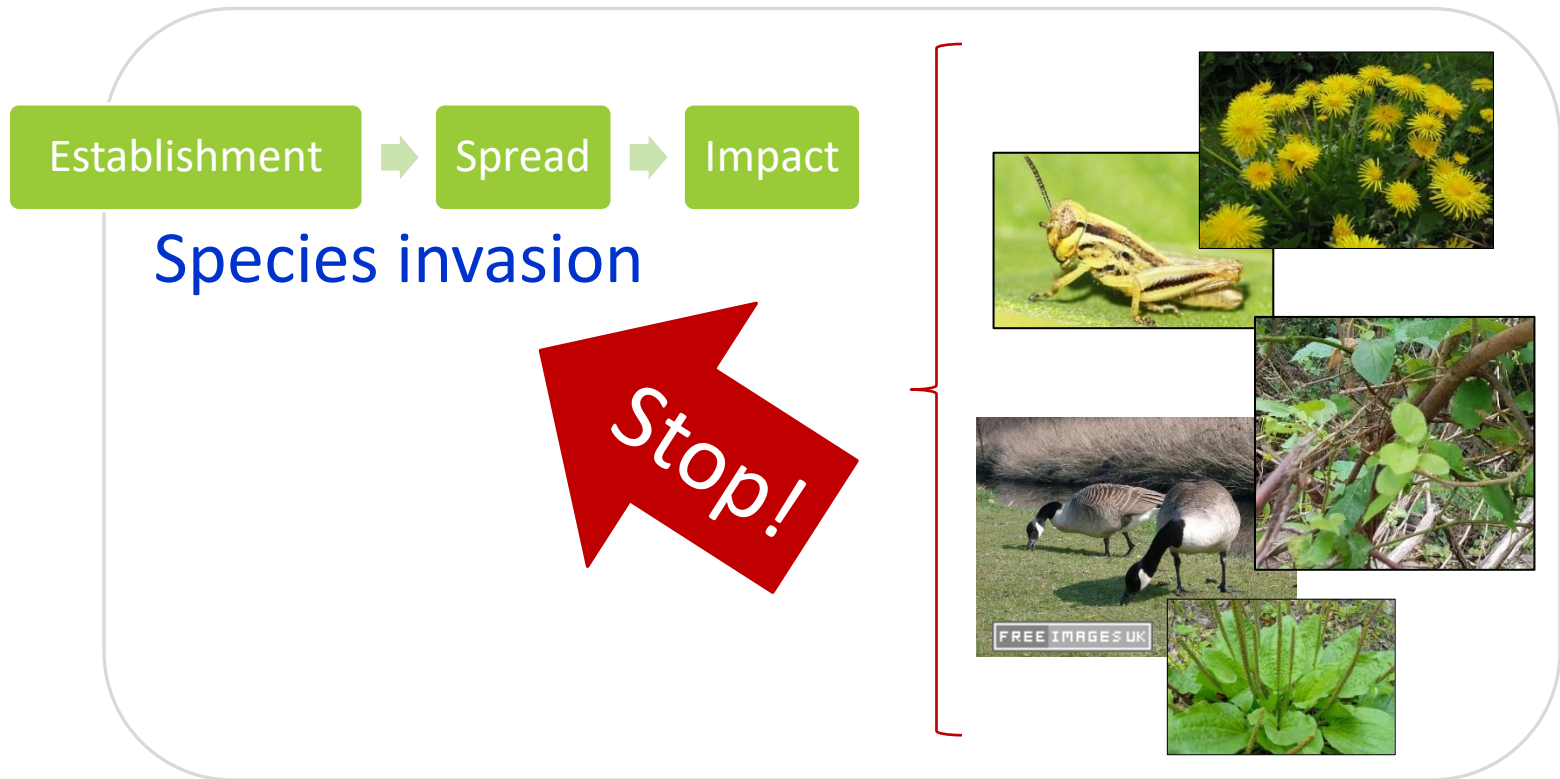
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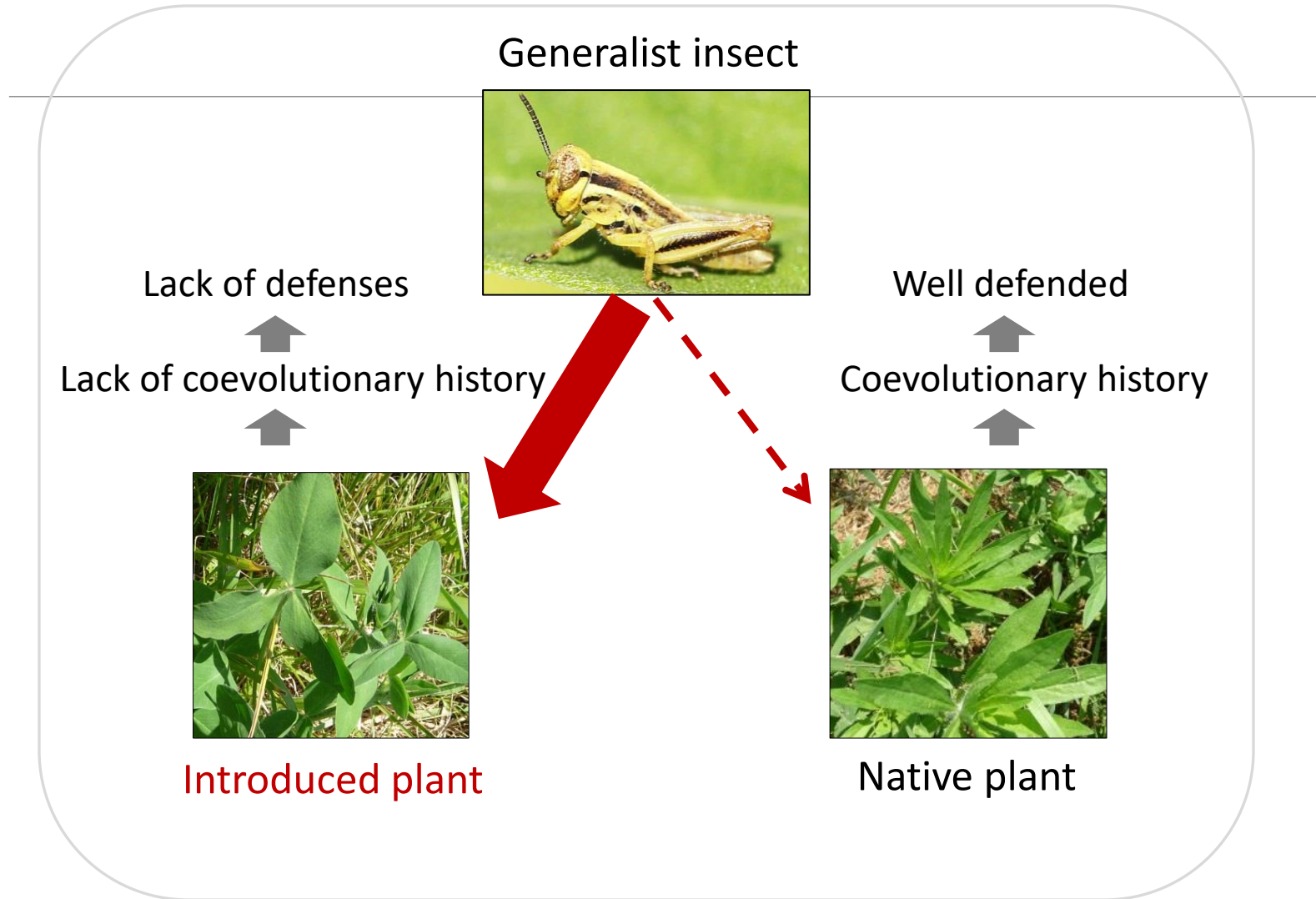
Why do introduced species fail to establish in a new range?

# Biotic resistance

- "the ability of resident species in a community **to reduce the success of exotic invasions**" (Levine et al., 2004) – i.e. competition, parasitism, herbivory, or predation, etc.



# Biotic Resistance Hypothesis



Native community

# Why do introduced species fail to establish in a new range?

---

## Novel species interactions



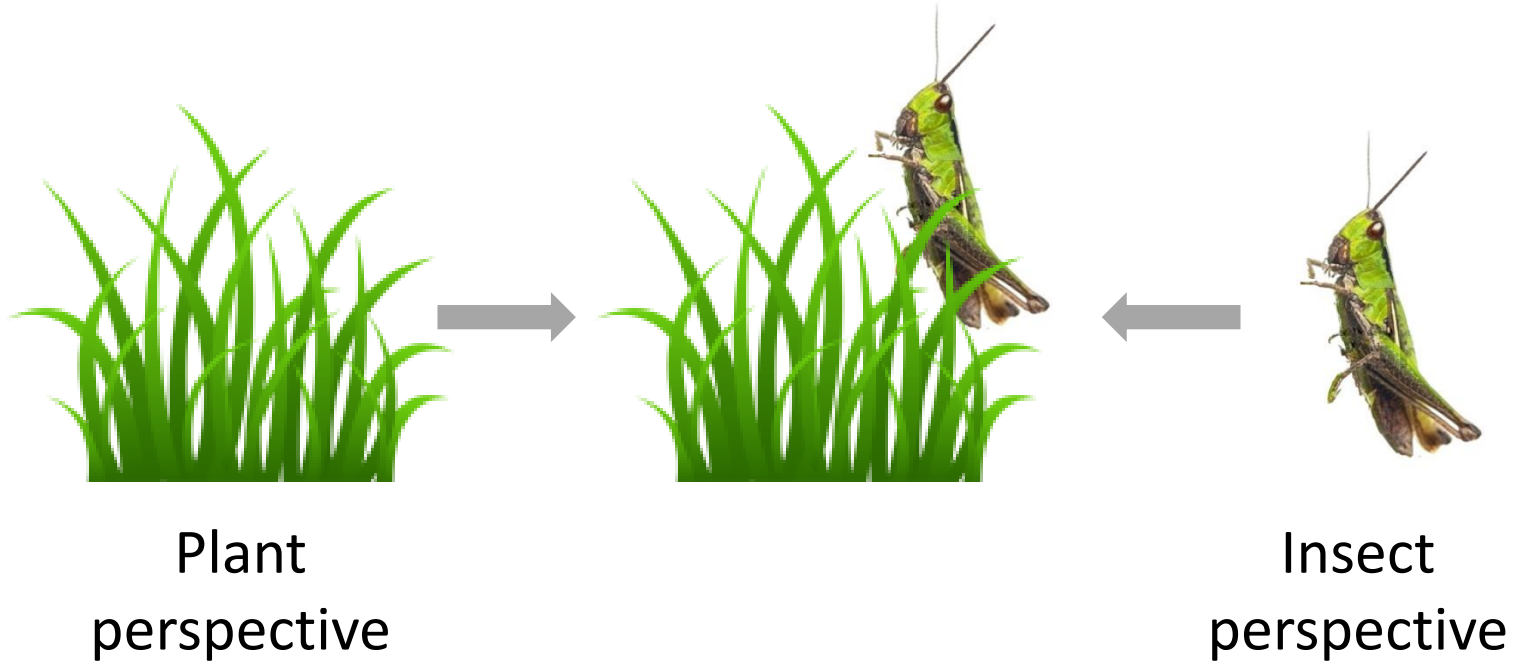
➤ How do insect herbivores respond to their novel host plants?



➤ How do plants respond to their novel insect herbivores?

# The Interaction between Generalist Insect Herbivores and Their Host Plants

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# Plant perspective

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Plant resistance to insect herbivory



Plant tolerance to insect herbivory



# Plant Resistance

- **The ability of a plant to decrease herbivore damage**

Price et al., 2011

- “A resistance trait is any plant character that influences the amount of damage a plant suffers”

Rausher, 1992



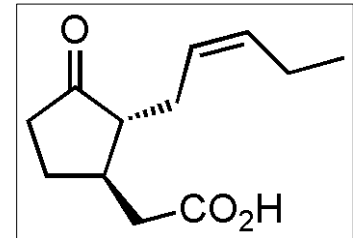
Wax



Spines



Trichomes



Jasmonic acid

- **Leaf damage** is one of the commonly used measurements for plant resistance
- Plants with more damage from herbivores are generally considered to have a lower level of resistance to herbivory

Mauricio 2000, Zou et al. 2008



# Plant Tolerance

- **The ability of a plant to maintain fitness while sustaining herbivore damage**

Price et al., 2011

- Physiological components of plant tolerance:  
growth rate, storage capacity, photosynthetic rates, nutrient uptake etc.

Rosenthal & Kotanen 1994

- **Plant compensatory growth in terms of aboveground plant biomass**  
is one of the fundamental and commonly used measurements for  
plant tolerance to herbivory, especially in grasslands

Rosenthal & Kotanen 1994; Atwood & Meyerson 2011;

Leis & Morrison 2011



- Estimating biomass should be **non-destructive**, accurate, and easy to implement

Redjadj et al. 2012





# Insect perspective

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*How do we know?*

➔ behavioral approach (feeding activity, consumption, assimilation)

➔ molecular approach (DNA barcoding of ingested plant material)



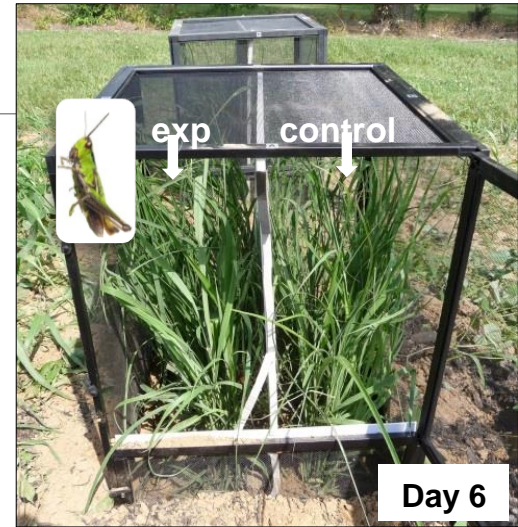
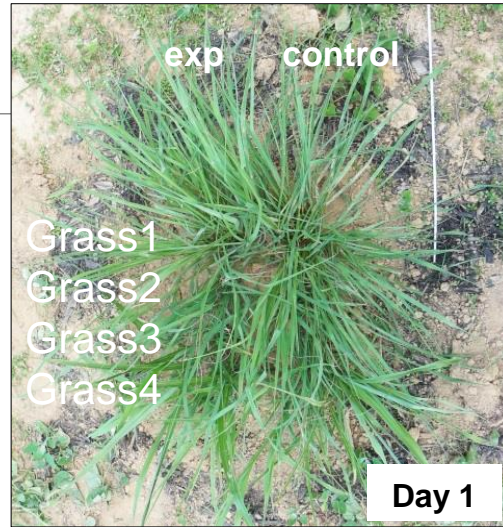
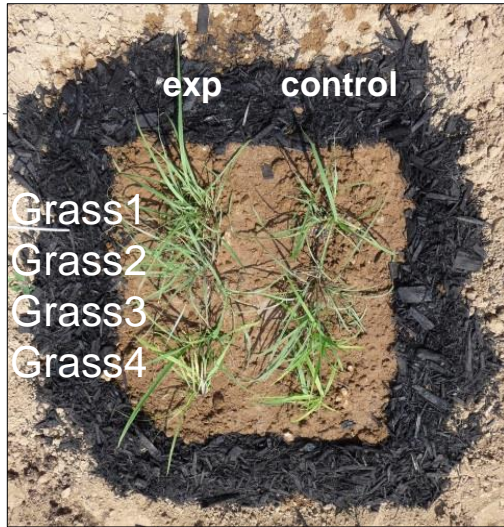
# Insect perspective

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behavioral approach (feeding activity, consumption, assimilation)

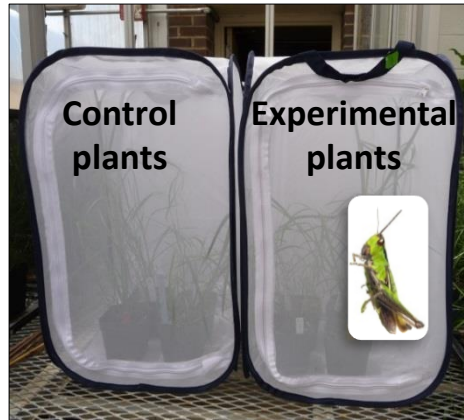
# Feeding Trials: Field



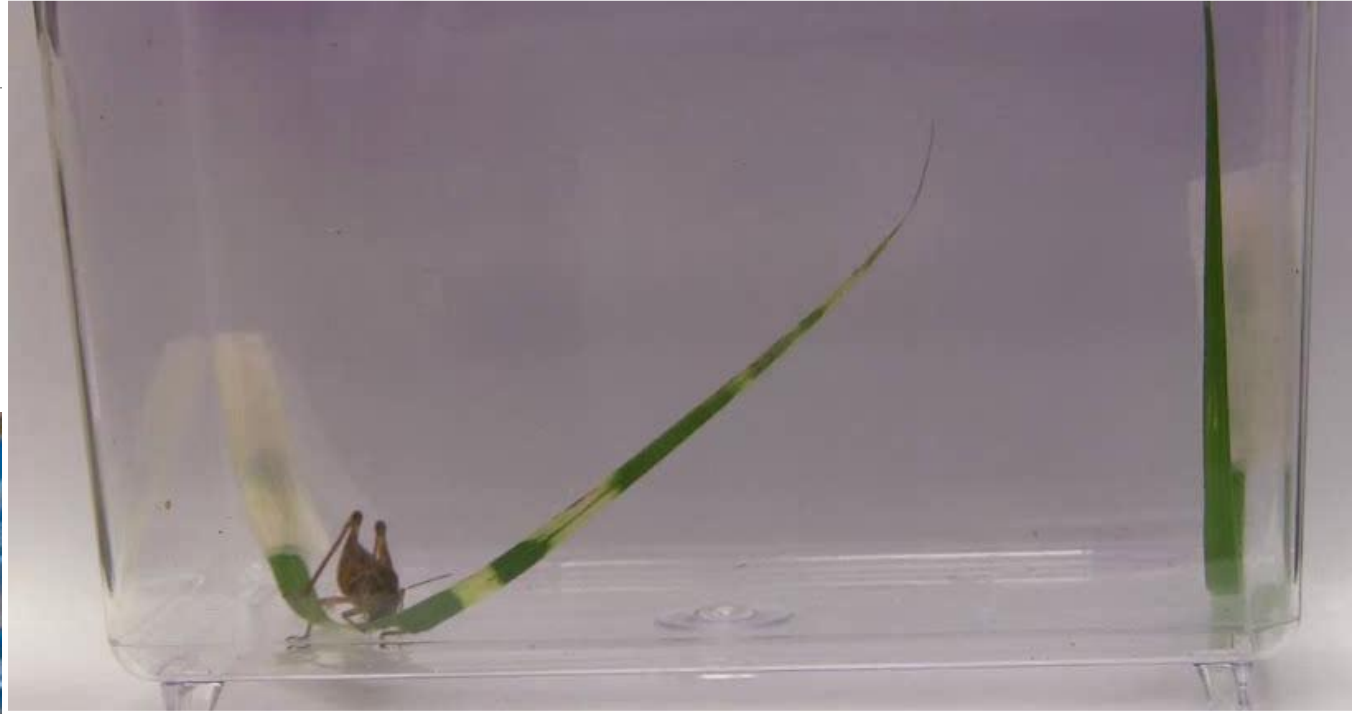
Plant growth / Grasshopper feeding

Plant regrowth  
Avanesyan and Culley (2017), *J. Torrey Soc.*

# Feeding Trials: UC Greenhouse



# Lab Assays (Leaves)





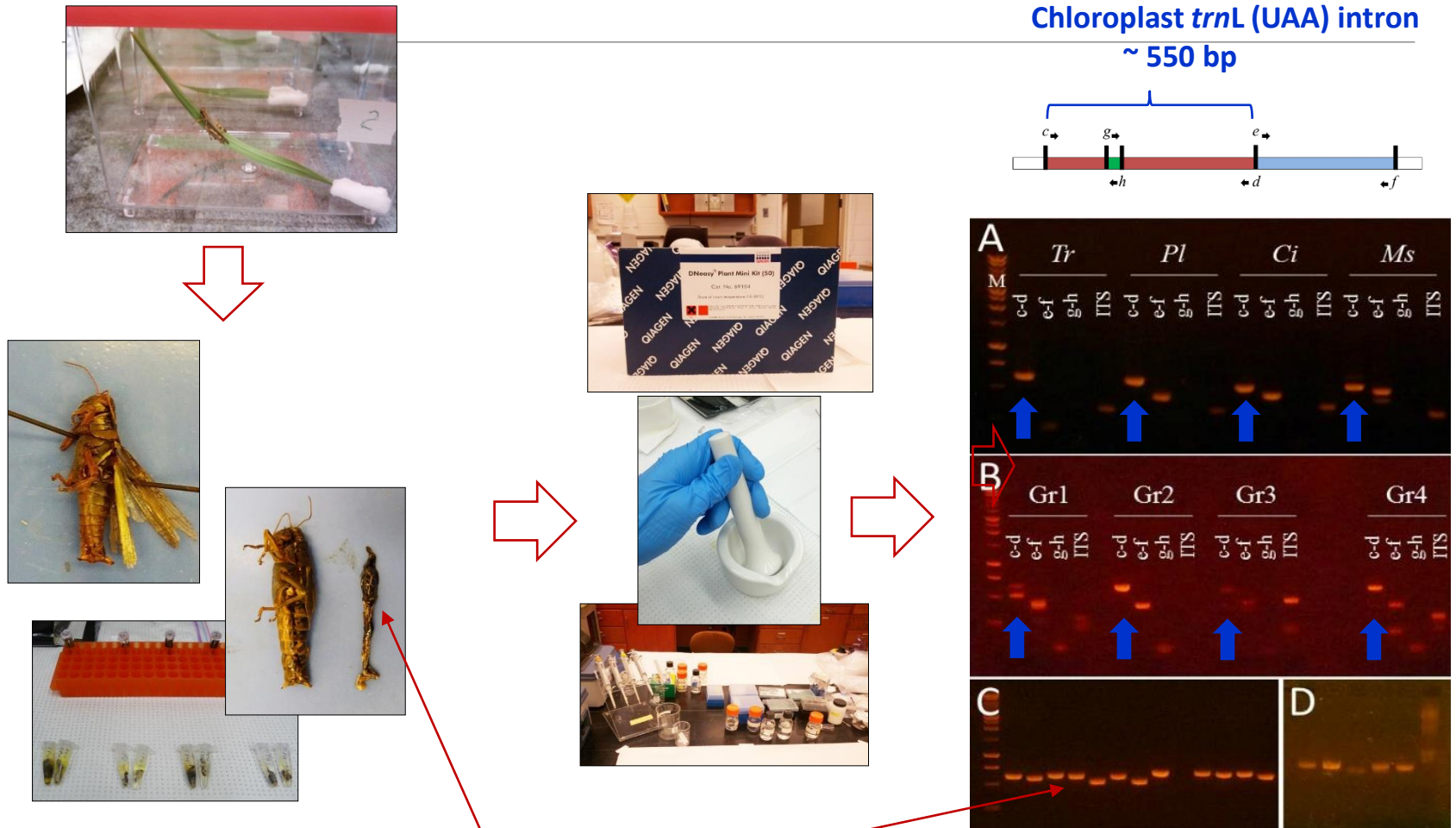
# Insect perspective

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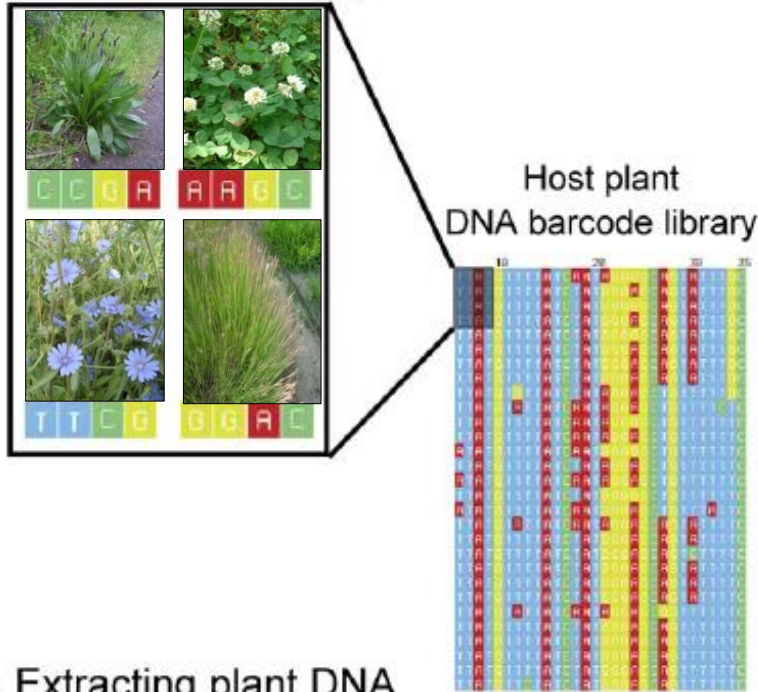
molecular approach (DNA barcoding of ingested plant material)

# Molecular Confirmation of Diet



# Host Plant Identification

## A. Assembling a host plant DNA barcode library

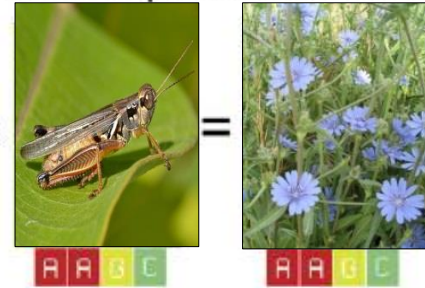


## B. Extracting plant DNA from insect herbivores



## C. Comparing extracted DNA with sequences in the DNA barcode library

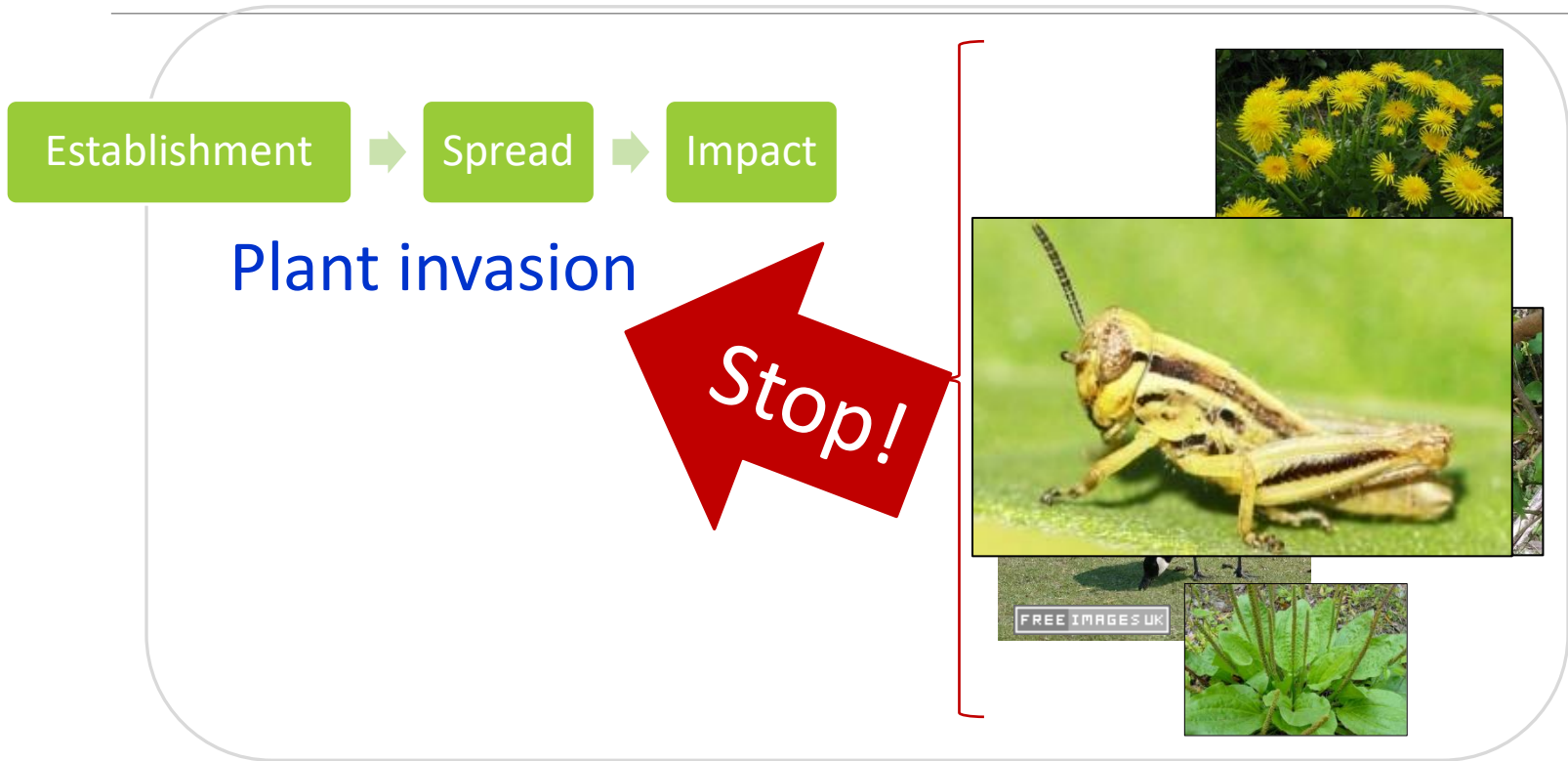
## D. Matching DNA sequences and host plant identification



- Plant ID
- Plant Origin



# Application to Biotic Resistance



Native community



# Quick Review

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- Can we determine insect feeding preferences for native vs. novel host plants? Yes/No
- Does biotic resistance refer to native species? Yes/No



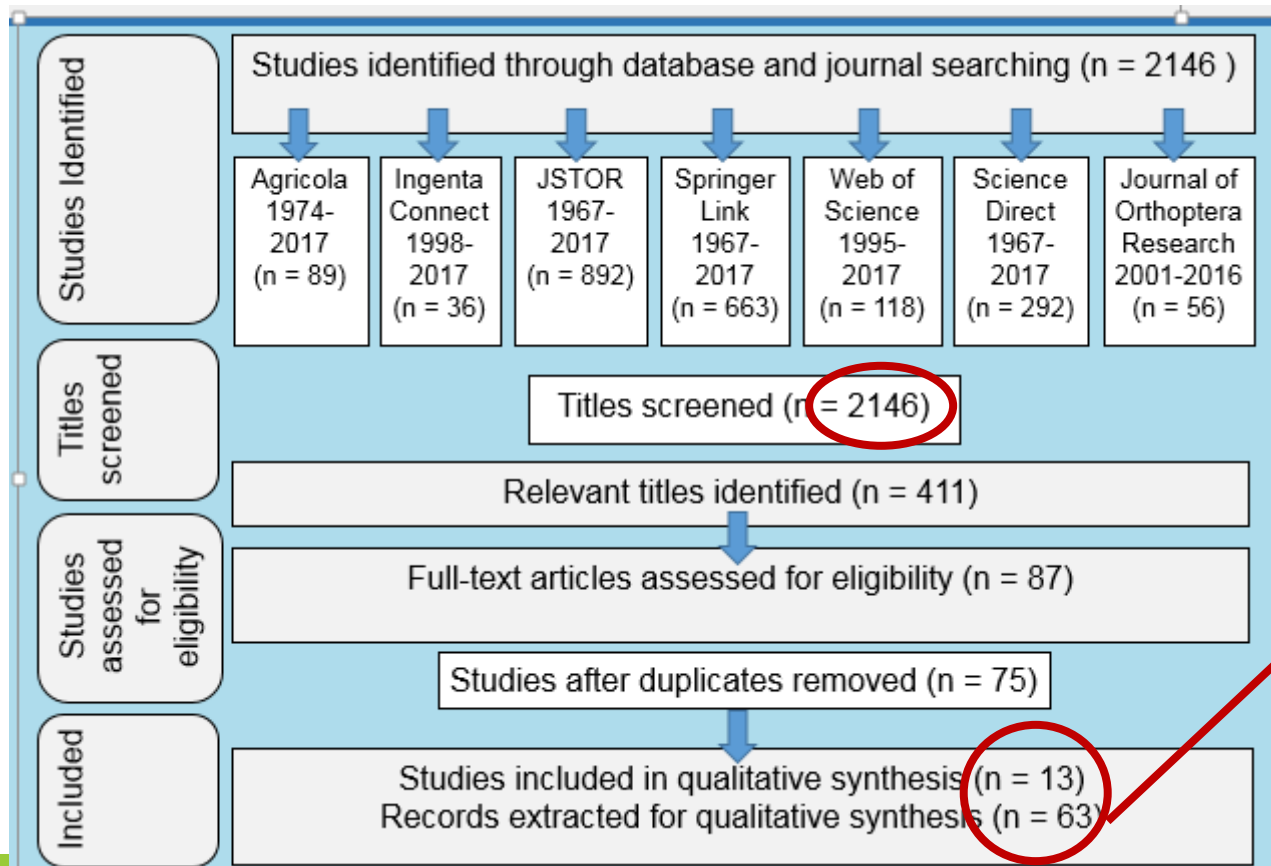
## Quick discussion (3 min)

---

→ If we know a feeding response for one insect species, does it mean all the insects in this genera/family, etc. have the same responses?

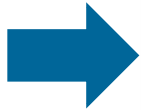
# Should I Eat or Should I Go?

## Acridid Grasshoppers and Their Novel Host Plants: Implications for Biotic Resistance



for 28 North-American grasshopper species

# Systematic Review and Meta-analysis



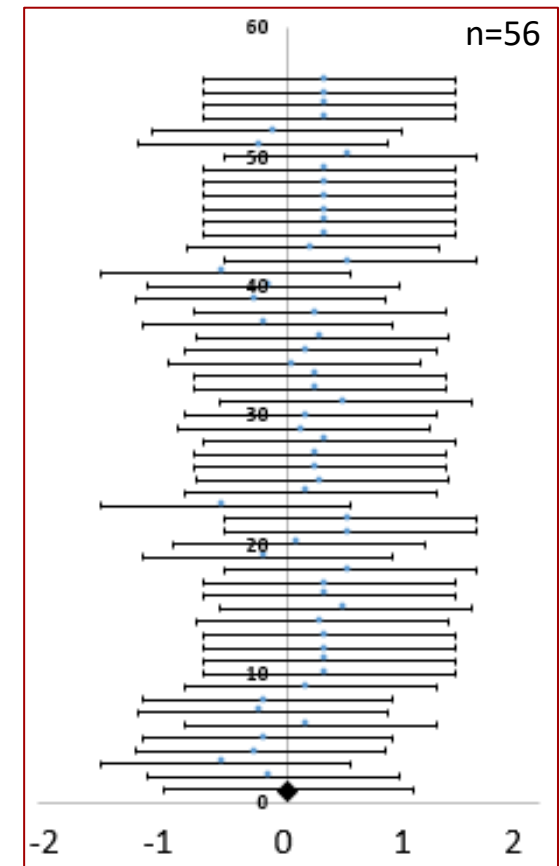
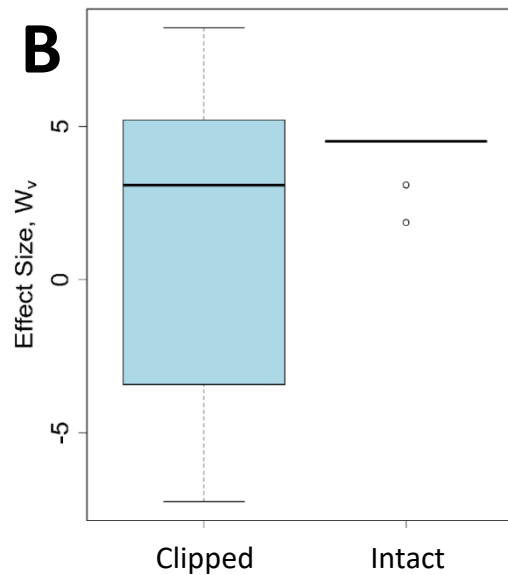
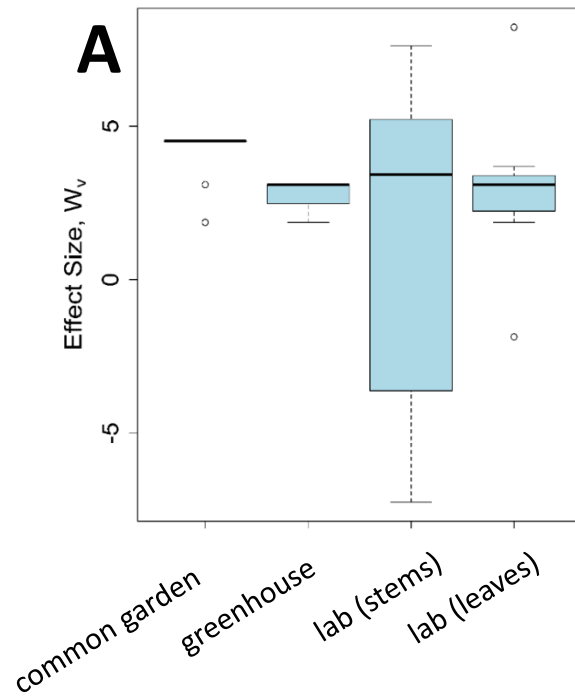
The authors used a very wide range of experimental conditions and measurements to assess grasshopper preferences

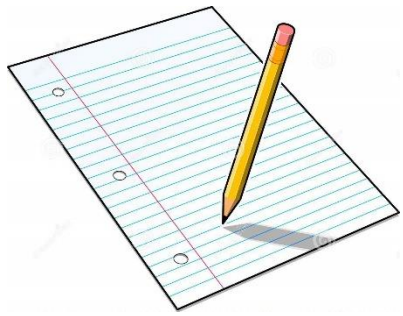
- **4 experimental environments:** common garden, greenhouse, lab (leaves), **lab (stems)**
- **3 types of feeding trials:** no-choice, choice (2 plants), **choice (plant mixture)**
- **2 types of plant material:** intact plants, **clipped plant parts**
- **Different stages:** **adults**, nymphs, mix
- **35 measurements** of feeding preferences!



# Discussion

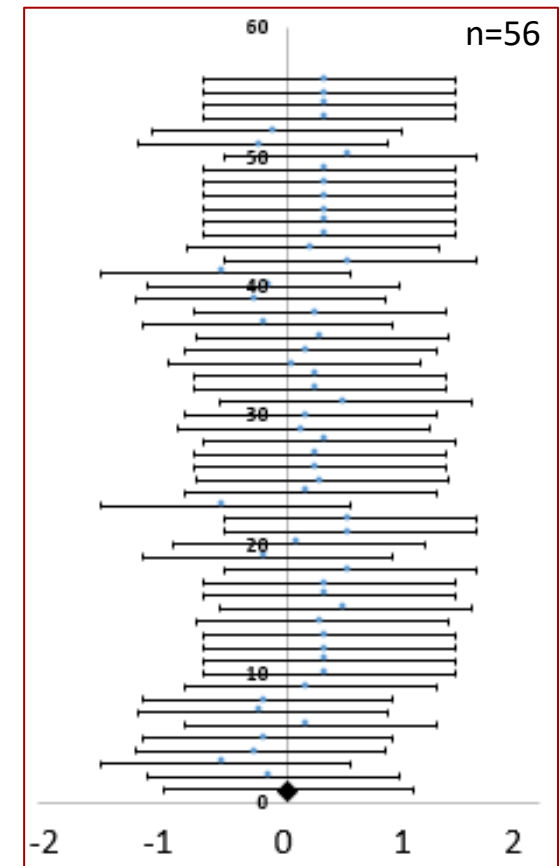
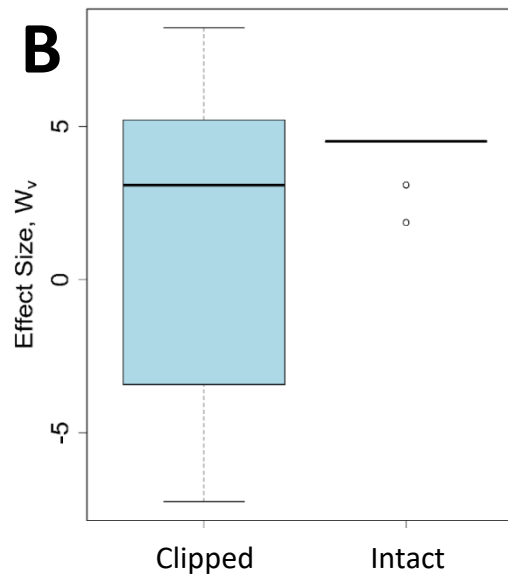
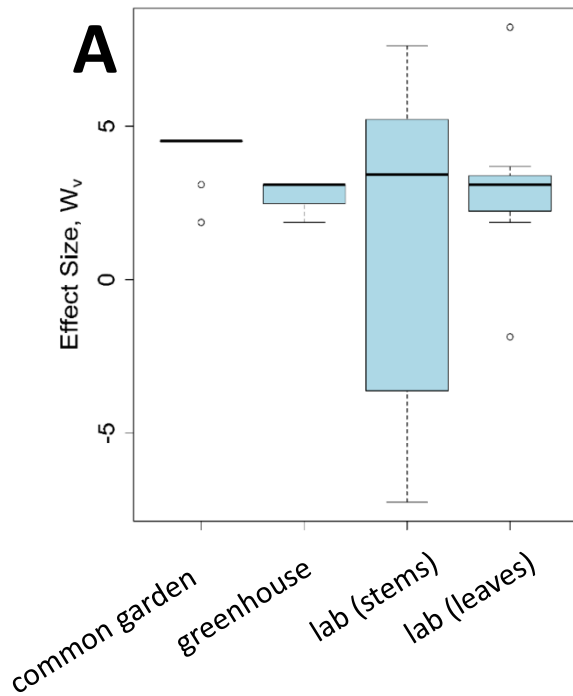
What do you see?





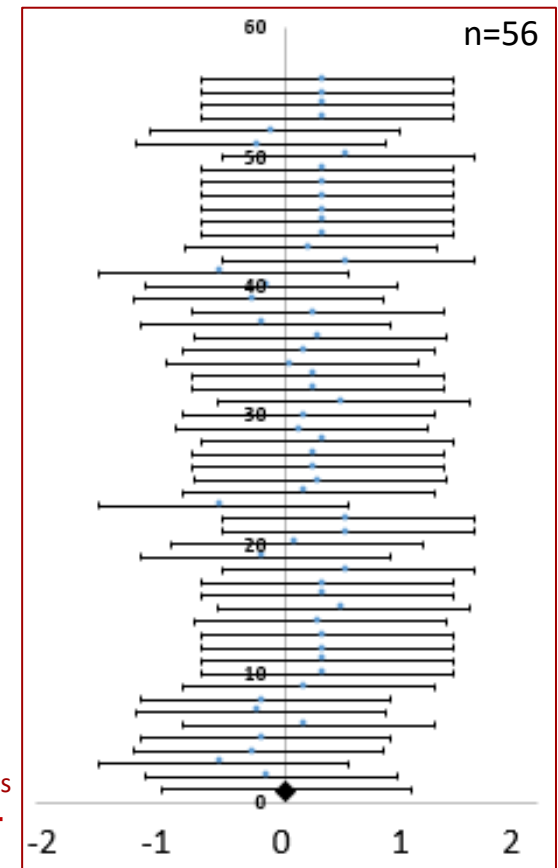
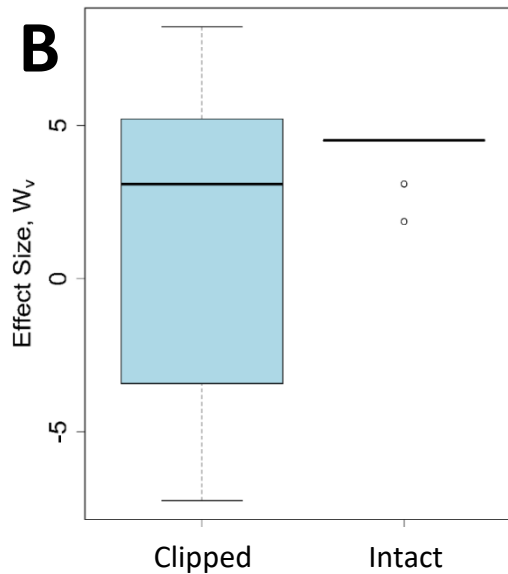
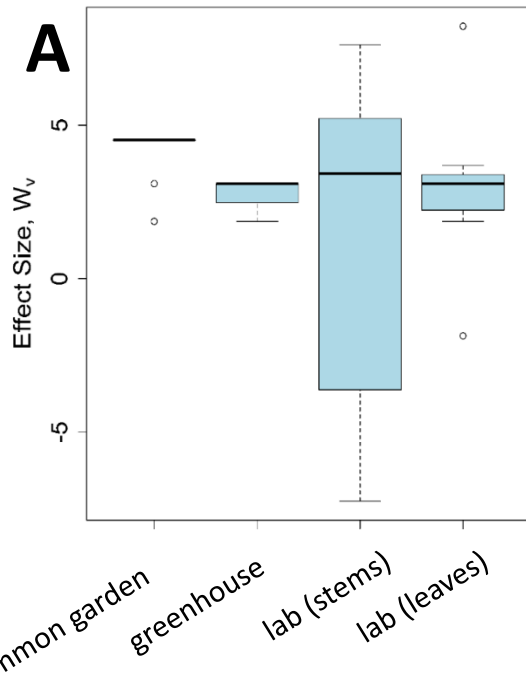
## Worksheet Part 4.

# What does it mean?



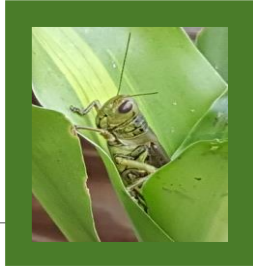


# What does it mean?

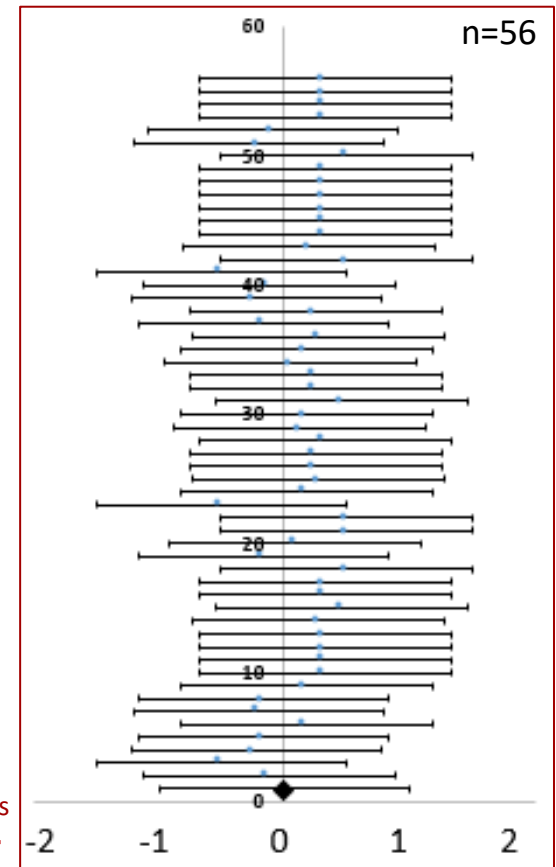
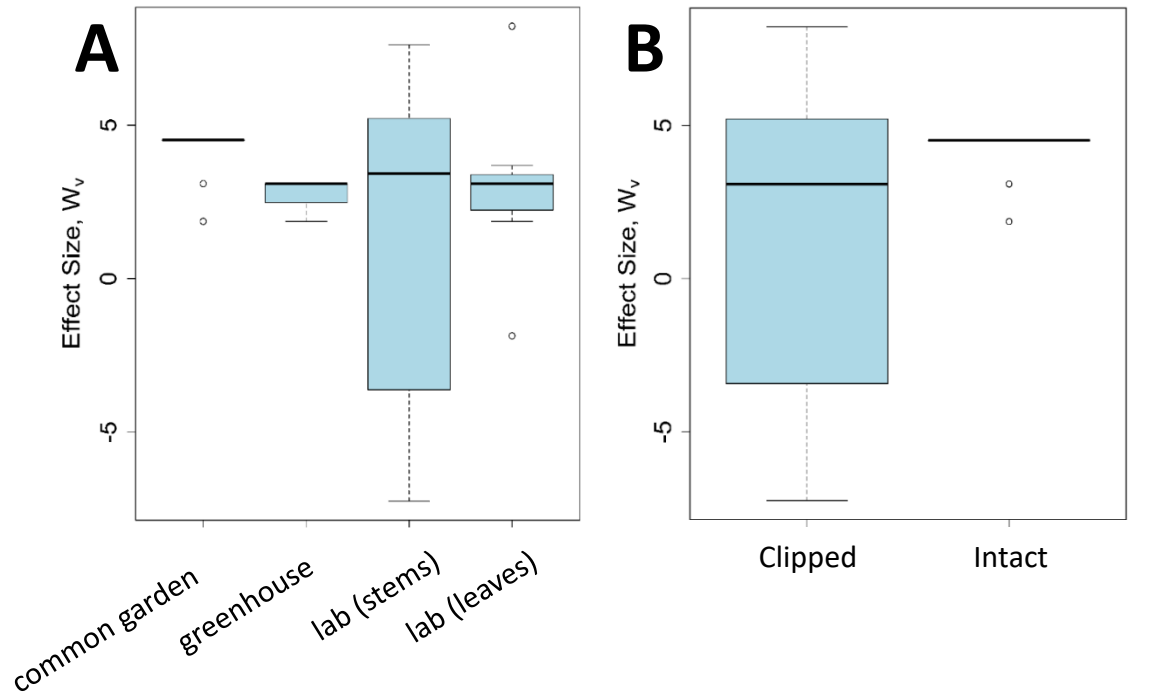


**Preference Metric** = 
$$\frac{n_{\text{most preferred exotic plant species}} - n_{\text{most preferred native plant species}}}{n_{\text{total plant species offered}}}$$





Acridid grasshoppers prefer to feed on introduced plants regardless the experimental conditions or plant material offered



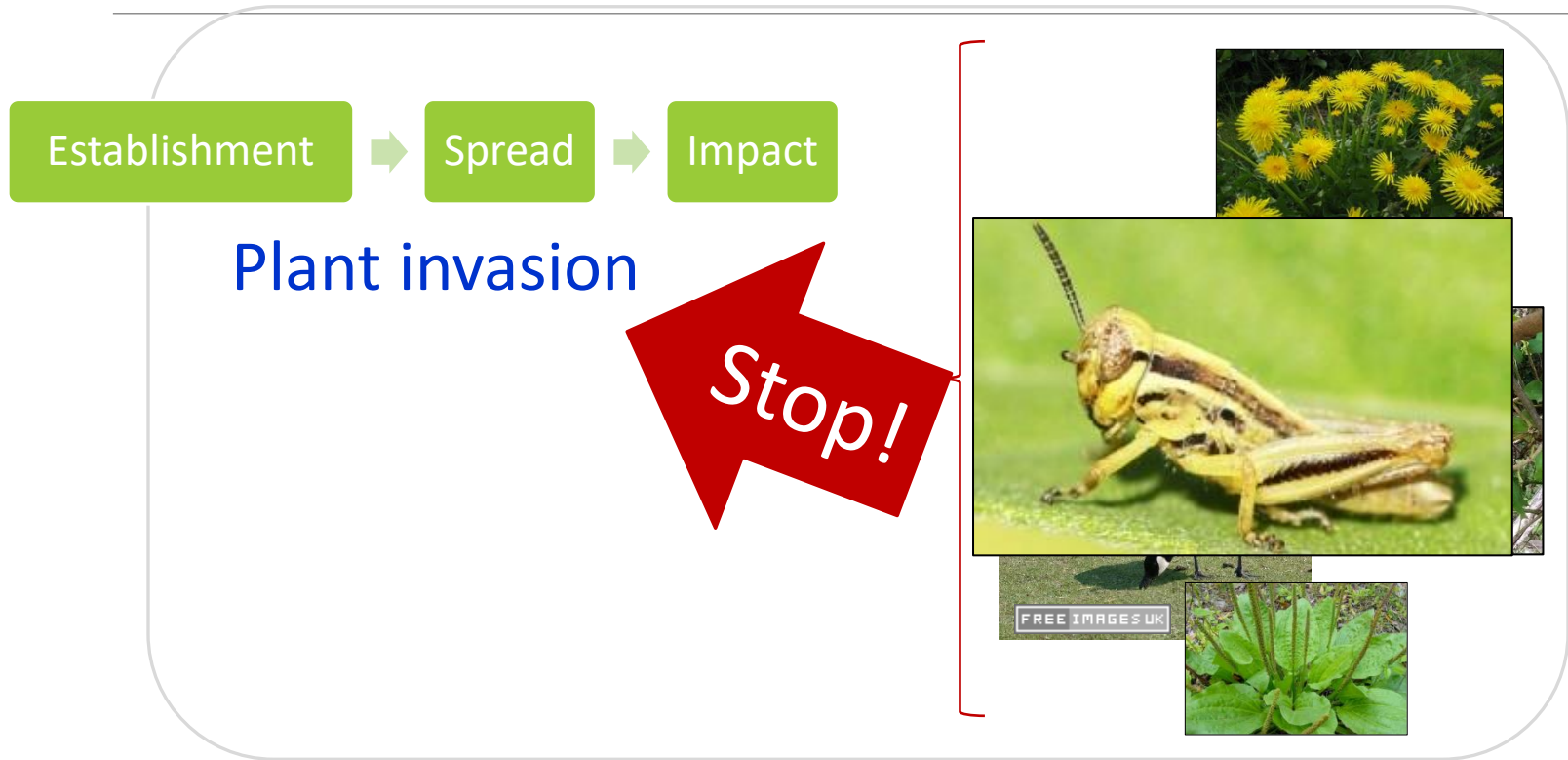
**Preference Metric** = 
$$\frac{n_{\text{most preferred exotic plant species}} - n_{\text{most preferred native plant species}}}{n_{\text{total plant species offered}}}$$



Most of the preferred plants are highly invasive

- ❖ 20 introduced plant species (out of 22) were reported as “the most preferred”
- ❖ 12 species showed high or middle invasive rank
- ❖ *Bromus inermis* (smooth brome) and *Schedonorus arundinaceus* (tall fescue) are among the most preferred (for 50% grasshopper species)

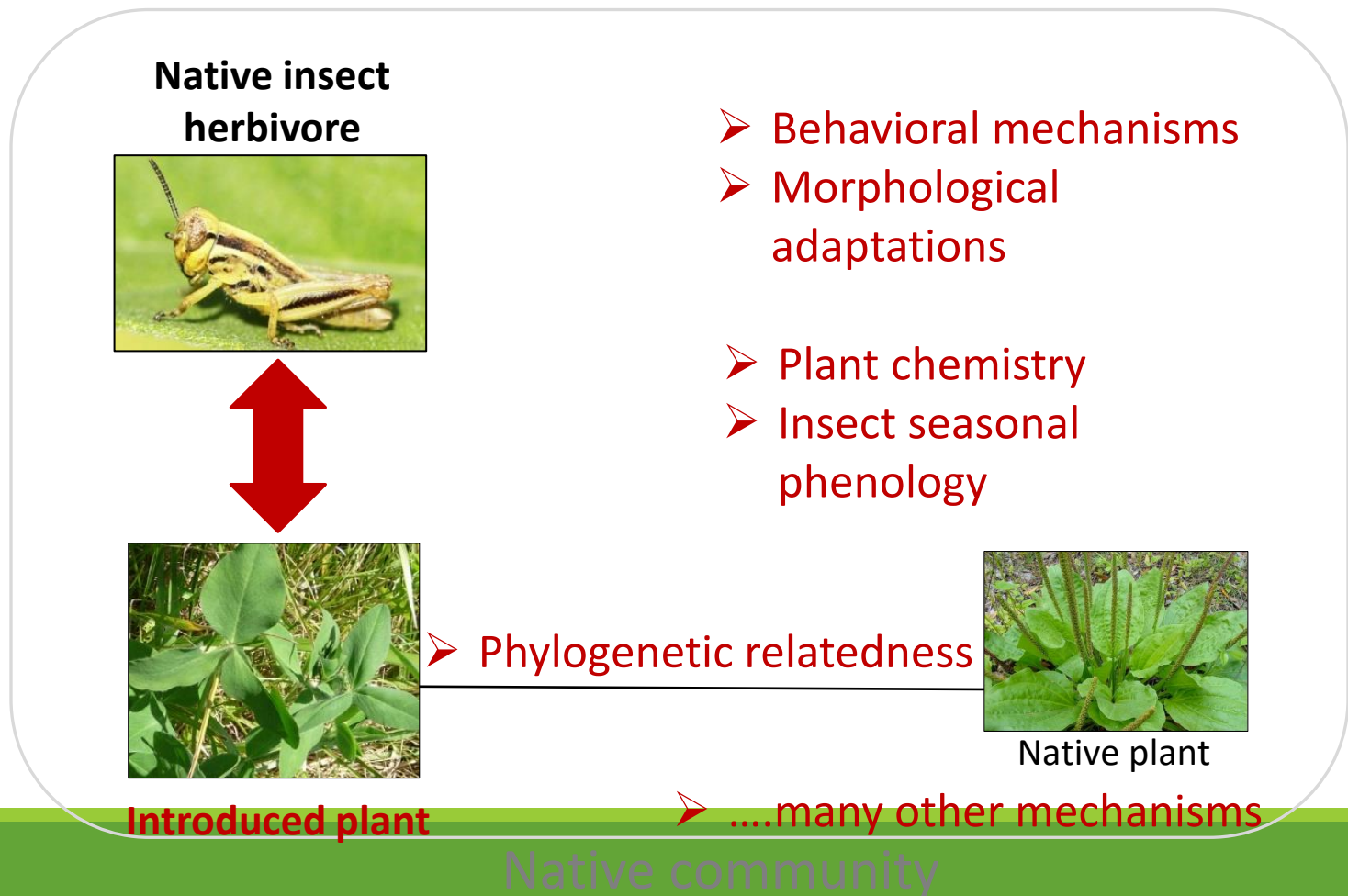
# Application to Biotic Resistance



Native community

# Summary

## Why do introduced species fail to establish in a new range?





# Quick Review

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- Can insect feeding behavior serve as a mechanism of biotic resistance? **Yes/No**
- Do all the introduced plants establish in the introduced range? **Yes/No**

# The End

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Please name at least one  
novel plant-insect association



# Image credits and resources

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USDA

Maryland Cooperative Extension

<https://www.nhbs.com/the-ecology-of-invasions-by-animals-and-plants-book>

<http://www.socialstudiesforkids.com>

<https://oceanservice.noaa.gov>

<https://www.ecori.org/natural-resources/2015/6/23/gypsy-moth-caterpillars-take-ri-by-storm>

Louis-Michel Nageleisen, Département de la Santé des Forêts, [Bugwood.org](http://Bugwood.org)

<http://bugoftheweek.com/blog/2017/4/29/good-bye-ash-trees-of-the-potomac-emerald-ash-borer-eab-iagrilus-planipennisi>