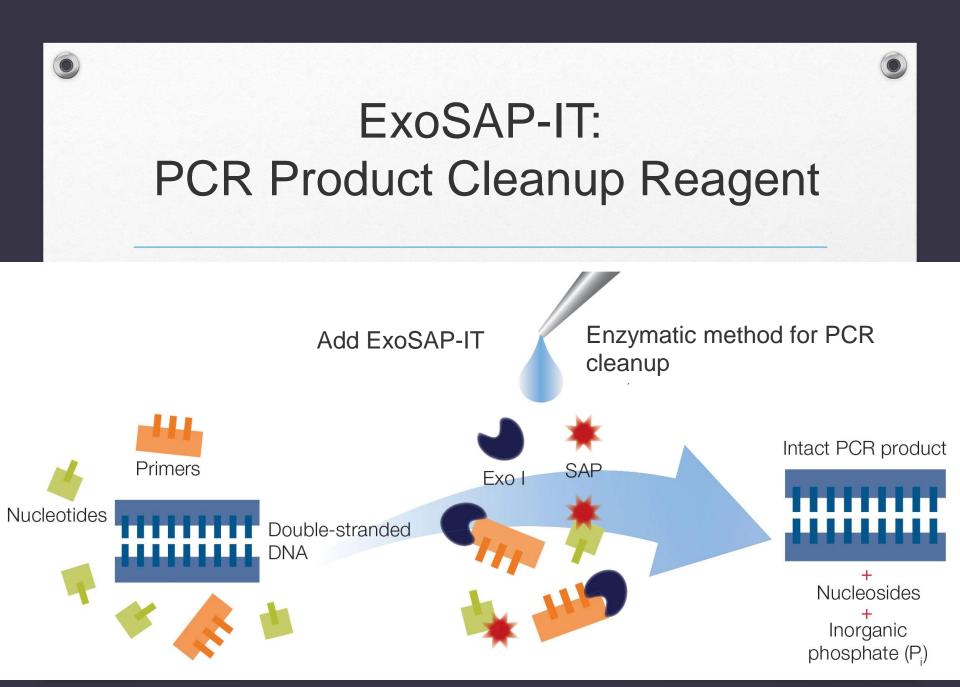




DNA Purification







Images: modified from https://www.thermofisher.com/us/en/home/life-science/sequencing/sanger-sequencing-kits-reagents/exosap-it-pcr-product-cleanup



PCR Equipment and Materials

Vortex



Racks





Pipette p20

Pipette tips





PCR machine

PCR tube strips



Marker



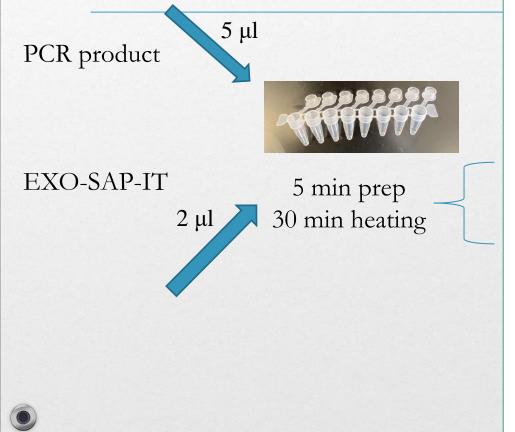
EXO-SAP-IT







DNA Purification Protocol



applied biosystems

QUICK REFERENCE

Thermo Fisher

SCIENTIFIC

ExoSAP-IT[™] PCR Product Cleanup

Brief Protocol

Catalog Number 78200, 78201, 78202, 78205, and 78250

Doc. Part No. 78200b Pub. No. MAN0016836 Rev. A.0 [02/2017]

MARNING! Read the Safety Data Sneets (SLOS) and Annual Columnia Colonia, and gloves. Safety Data Sneets (SDSs) are available WARNING! Read the Safety Data Sheets (SDSs) and follow the from thermofisher.com/support.

Product description

ExoSAP-IT" reagent treats PCR products ranging in size from less than 100 bp to over 20 kb with absolutely no sample loss by removing unused primers and nucleotides. Add ExoSAP-IT" reagent directly to the reaction products following PCR. ExoSAP-IT" PCR Product Cleanup is active in commonly used PCR buffers, so no buffer exchange is required. After treatment. ExoSAP-IT²⁰ reagent is inactivated by heating to 80°C for 15 minutes. The treated PCR products are now ready for subsequent analysis in applications that require DNA to be free of excess primers and nucleotides.

PCR cleanup protocol

- Note: Store ExoSAP-IT[™] reagent at -20°C in a non-frost-free freezer. Remove ExoSAP-IT[™] reagent from -20°C freezer and keep on ice throughout this procedure.
- 2. Mix 5 µL of a post-PCR reaction product with 2 µL of ExoSAP-IT" reagent for a combined 7 µL reaction volume.
- When treating PCR product volumes greater than 5 µL, simply increase the amount of ExoSAP-IT¹⁸ reagent proportionally. 3. Incubate at 37°C for 15 minutes to degrade remaining primers and nucleotides.
- Incubate at 80°C for 15 minutes to inactivate ExoSAP-IT^{**} reager alina
- The PCR product is now ready for use in DNA sequencing, SNI analyses, or other primer-extension applications. Treated PCR products may be stored at -20°C until required.

ExoSAP-IT[™] Cleanup product overview

ExoSAP-IT Cleanup

Customer and technical support Visit thermofisher.com/support for the latest in services and support,

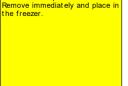
including: · Worldwide contact telephone numbers

- · Product support, including:
- Product FAQs
- Software, patches, and updates
- Training for many applications and instruments · Order and web support
- · Product documentation, including:
- User guides, manuals, and protocols Certificates of Analysis
- Safety Data Sheets (SDSs; also known as MSDSs)
- Note: For SDSs for reagents and chemicals from other manufacturers, contact the manufacturer.

Limited product warranty

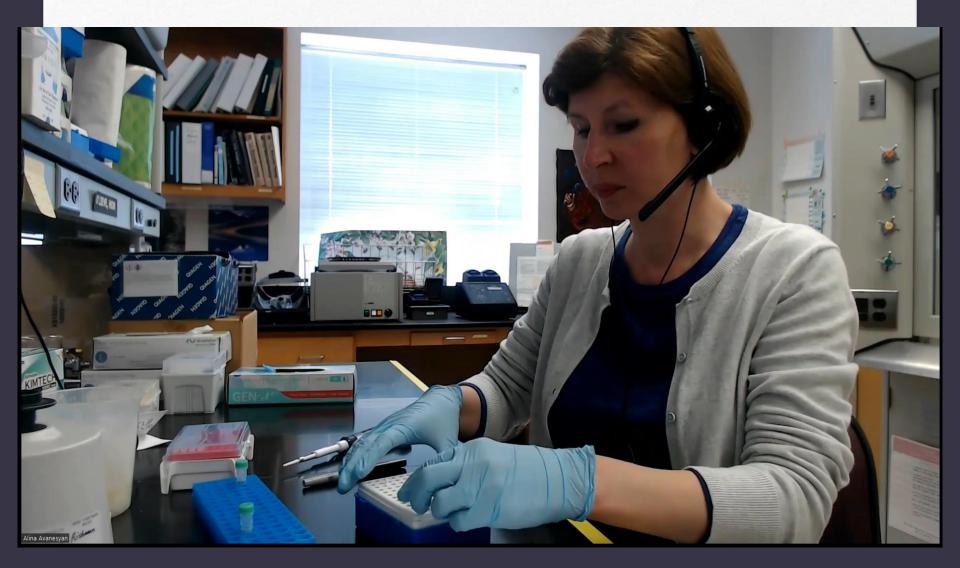
Life Technologies Corporation and/or its affiliate(s) warrant their products as set forth in the Life Technologies' General Terms and Conditions of Sale found on Life Technologies' website at www.thermofisher.com/us/en/home/global/terms-andconditions.html. If you have any questions, please contact Life Technologies at www.thermofisher.com/support.

Use PCR protocol "80" in the PCR machine to conduct both ncubation steps.





Demo 1: DNA Purification Prep



Before you go...

- After completing the prep EXO-SAP-IT and PCR product leftovers should be placed back to the freezer
- Once the heating rounds are done the PCR tubes should be immediately removed from the PCR machine.
- Then you can either store the purified PCR products at -20°C until you need to submit them for sequencing, or you can proceed immediately to sample preparation for sequencing.





Image and video credits

- Videos: DNA purification prep was demonstrated by Alina Avanesyan; recording and editing were done by Alina Avanesyan
- Photos: preparing and editing by Anya Wilkinson and Alina Avanesyan
- DNA purification protocol: modified by Alina Avanesyan; original manufacturer's protocol is available at Thermo Fisher Scientific (https://www.thermofisher.com/)
- Videos were recorded and photos were taken in Dr. David Hawthorne's lab: 4172 Plant Science Building, Department of Entomology, University of Maryland, College Park, MD

Acknowledgements

• We thank Dr. David Hawthorne (Department of Entomology, University of Maryland) for providing lab equipment and lab space for our DNA barcoding work; for providing lab space to take the photos and record the videos needed for developing this course; and for continuous support and encouragement!