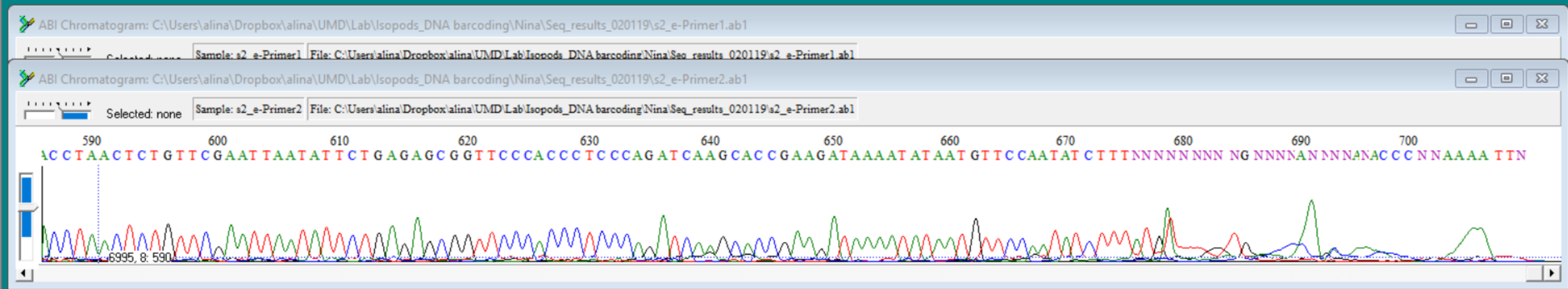


Open both sequences for primers 1 and 2; put trace file and sequence file to see them together for one sample



DNA sequence from C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\s2_e-Primer2.ab1
1 total sequences

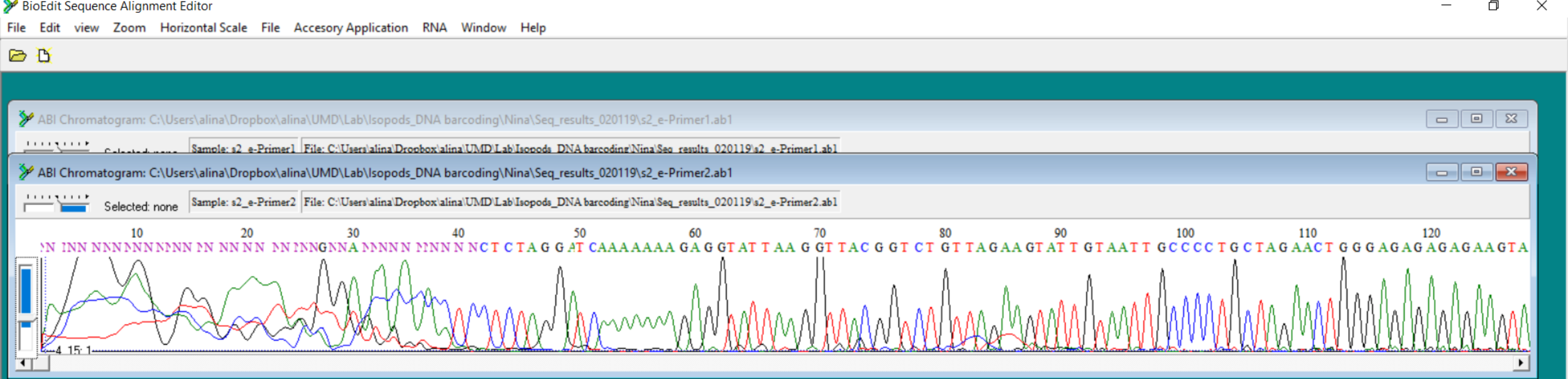
Courier New 11 B

Mode: Edit Overwrite Selection: null Position: Sequence Mask: None Numbering Mask: None Start ruler at: 1

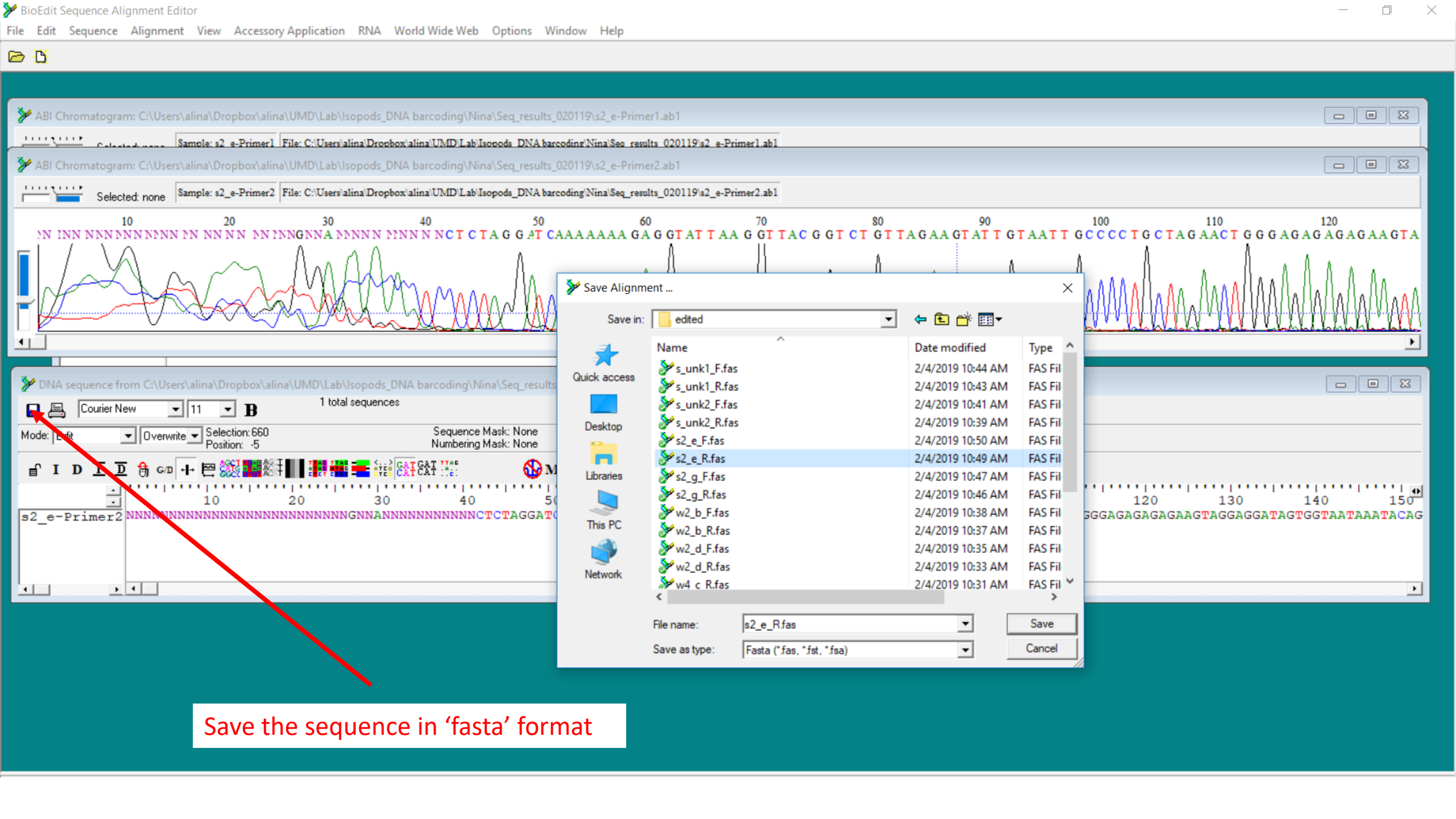
Scroll speed slow fast

560 570 580 590 600 610 620 630 640 650 660 670 680 690 700
s2_e-Primer2 GATCATTTCCAAATAAATCTCCAGGTTGACCTAACTCTGTTTCGAATTAATATTTCTGAGAGCGGTTCCACCCCTCCAGATCAAGCACCGAAGATAAAATATAATGTTCCAATATCTTTNNNNNNNNNGNNNNANNNANACCNNAAAAATTN

Scroll both files to the end; click on the sequence file; put the cursor at the position you want to delete; change mode to 'Edit'; click on 'delete'



Scroll both files to the beginning; repeat the editing using 'backspace'



ABI Chromatogram: C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\s2_e-Primer1.ab1

Sample: s2_e-Primer1 File: C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\s2_e-Primer1.ab1

ABI Chromatogram: C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\s2_e-Primer2.ab1

Sample: s2_e-Primer2 File: C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\s2_e-Primer2.ab1

10 20 30 40 50 60 70 80 90 100 110 120

IN INN NNNNNNNN NN NN NN NN INNGNA NNNN NNN NCTCTAG GATCAAAAAA GAGGTAT TAA GGT TAC GGTCT GTTAGAAGTAT T GTAATT GCCCCTGCTAGA ACTGGGAGAGAGAGAAGTA

DNA sequence from C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results

1 total sequences

Mode: Edit Overwrite Selection: 660 Position: -5 Sequence Mask: None Numbering Mask: None

s2_e-Primer2 NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNGNNA NNNNNNNNNNNNNCTCTAGGATC

Save Alignment ...

Save in: edited

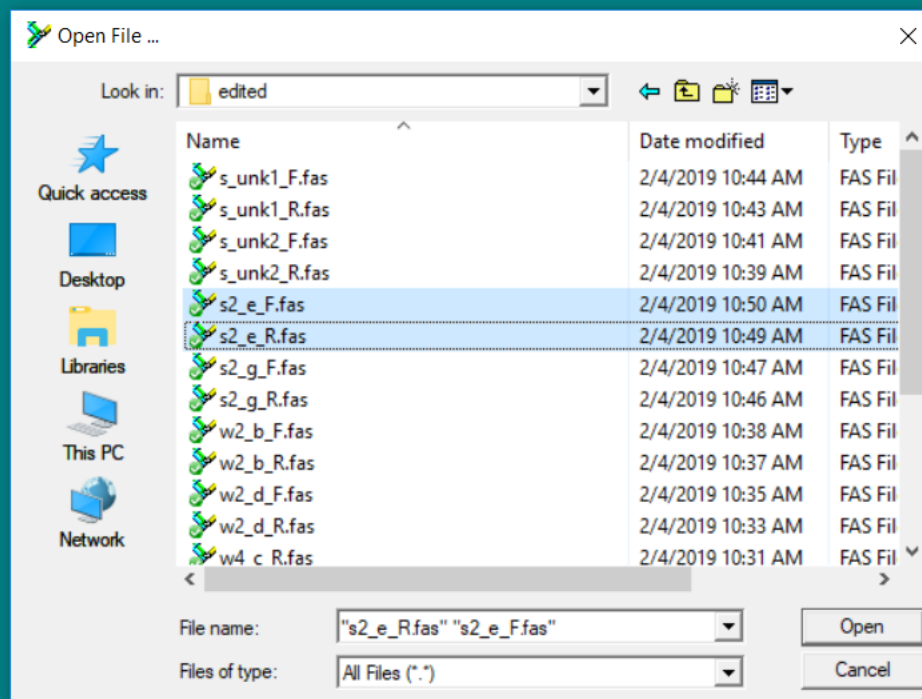
Name	Date modified	Type
s_unk1_F.fas	2/4/2019 10:44 AM	FAS Fil
s_unk1_R.fas	2/4/2019 10:43 AM	FAS Fil
s_unk2_F.fas	2/4/2019 10:41 AM	FAS Fil
s_unk2_R.fas	2/4/2019 10:39 AM	FAS Fil
s2_e_F.fas	2/4/2019 10:50 AM	FAS Fil
s2_e_R.fas	2/4/2019 10:49 AM	FAS Fil
s2_g_F.fas	2/4/2019 10:47 AM	FAS Fil
s2_g_R.fas	2/4/2019 10:46 AM	FAS Fil
w2_b_F.fas	2/4/2019 10:38 AM	FAS Fil
w2_b_R.fas	2/4/2019 10:37 AM	FAS Fil
w2_d_F.fas	2/4/2019 10:35 AM	FAS Fil
w2_d_R.fas	2/4/2019 10:33 AM	FAS Fil
w4 c R.fas	2/4/2019 10:31 AM	FAS Fil

File name: s2_e_R.fas Save

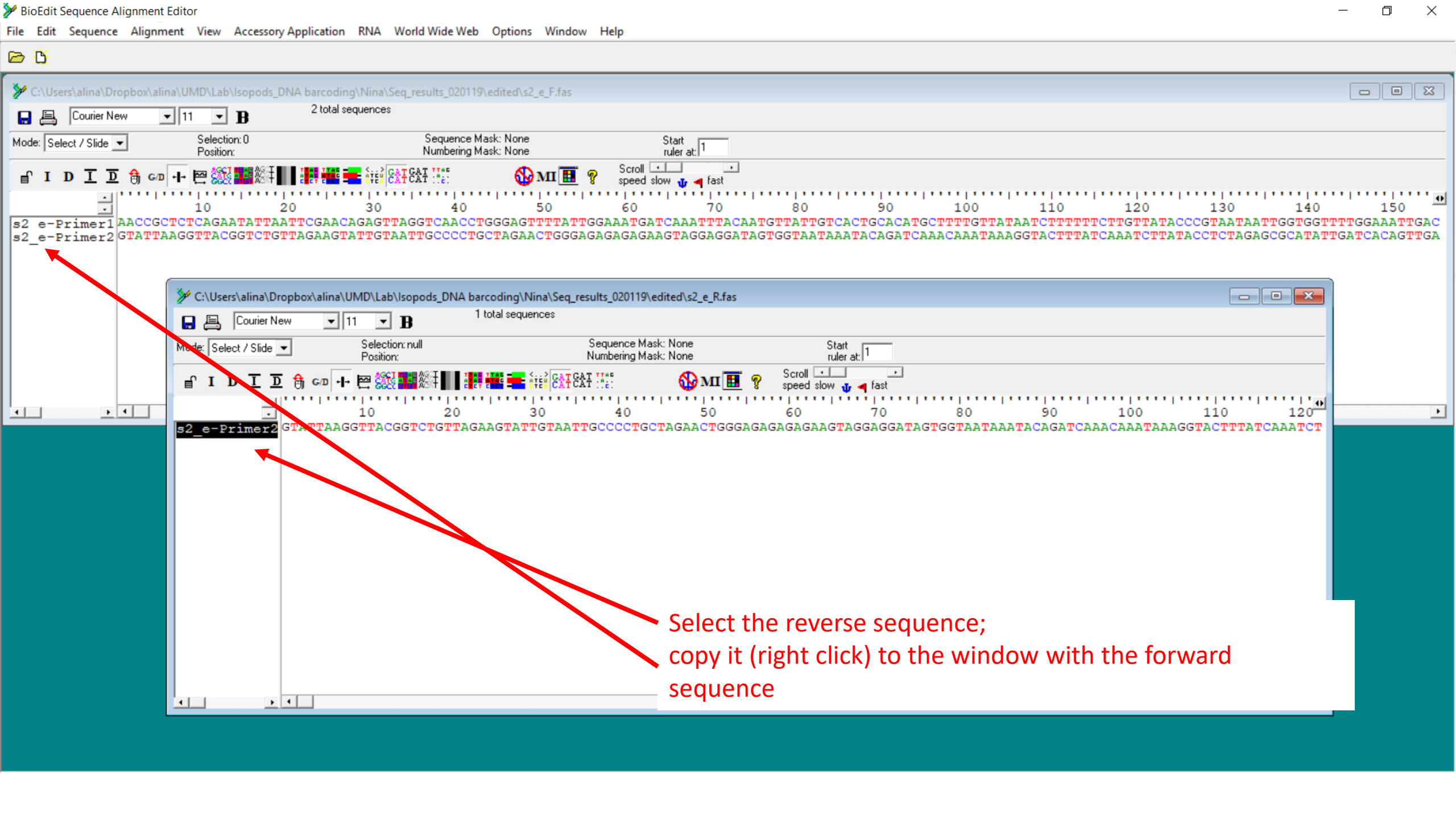
Save as type: Fasta (*.fas, *.fst, *.fsa) Cancel

Save the sequence in 'fasta' format

Do the editing for the other sequence; then close all the windows



Then, open both forward and reverse edited sequences



Select the reverse sequence;
copy it (right click) to the window with the forward
sequence

- New Sequence
- Edit Sequence
- Edit all selected
- Select Positions
- Open at cursor position
- Extract Positions
- Go to Pubmed references
- Overwrite/Retrieve sequences by gi number in title by HTTP to GenBank
- Overwrite/Retrieve Genbank data by gi number in title by HTTP to GenBank (ignore sequence)
- True positions from alignment positions
- Phylogeny / Taxonomy >
- Filter out sequences containing certain characters >
- Rename >
- Sort >
- PCR Primers / oligos >
- Pairwise alignment >
- Similarity Matrix (for pairwise alignments and shading) >
- Features >
- Sequence groups (or families) >
- Edit Mode >
- Mask >
- Toggle Color
- Gaps >
- Manipulations >
- Nucleic Acid >**
- Protein >
- Translate or Reverse-Translate (permanent)
- Translate in selected frame (permanent)
- Toggle Translation Ctrl+G
- Toggle translation in selected frame
- Toggle translation of CDS annotations
- Dot Plot (pairwise comparison)

C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\edited\s2_e_F.fas
2 total sequences

Courier New 11 B
Mode: Select / Slide Selection: 0 Position: 66
Sequence Mask: None Numbering Mask: None Start ruler at:

s2_e-Primer1 AACCGCTCTCAGAAATTTAATTCGAACAGAGTTAGGTC AACCTGGGAGTTTATTGGAAATGATCAAAT
s2_e-Primer2 GTATTAAGGTTACGGTCTGTTAGAAGTATTGTAATTGCCCTGCTAGAACTGGGAGAGAGAGAGTAGG

- Nucleotide Composition
- Base composition and mass export (monoisotopic)
- Base composition and mass export with average masses
- Complement
- Reverse Complement Shift+Ctrl+R**
- DNA->RNA
- RNA->DNA
- Translate >
- Find next ORF
- Create Plasmid from Sequence
- Gap beginning to minimize stop codons in reading frame 1
- Restriction Map
- Sorted Six-Frame Translation
- Unsorted Six-Frame Translation
- Find ORFs from a list of positions

Select the reverse sequence;
Do reverse compliment



Add / Modify / Remove an Accessory Application

ClustalW Multiple alignment

BLAST

CAP contig assembly program

ClustalW Example Application

DNADist ---> Neighbor phylogenetic tree

DNADist DNA distance matrix

DNAMl DNA Maximum Likelihood program

DNAMlk DNA Maximum Likelihood program with molecular clock

DNAPars DNA parsimony method

FastDNAMl DNA maximum likelihood

Fitch -- Fitch-Margoliash and Least-Squares Distance Methods

Kitsch -- Fitch-Margoliash and Least Squares Methods with Evolutionary Clock

NEIGHBOR -- Neighbor-Joining and UPGMA methods

ProML Protein Maximum Likelihood program

Protdist ---> Fitch phylogenetic tree

Protdist ---> Neighbor phylogenetic tree

Protdist protein distance matrix

Protpars protein parsimony method

C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\edited\s2_e_F.fas

2 total sequences

Mode: Select / Slide Selection: 0 Position: Sequence Mask: None Numbering Mask: None Start ruler at:

I D I D G-D + -

s2_e-Primer1 AACCGCTCTCAGAAATTTAATTCGAACAGAGTTAGGTC AACCTGGGAGTTTATTGGAAATGATCAAAI

s2_e-Primer2 TTATATTTTATCTTCGGTGCTTGATCTGGGAGGGTGGGAACCGCTCTCAGAAATTTAATTCGAACAGAG

Then select both sequences;
Do alignment

ClustalW Options

ClustalW Multiple alignment

Reference:
Thompson, J.D., Higgins, D.G. and Gibson, T.J. (1994)
CLUSTAL W: improving the sensitivity of progressive multiple
sequence alignment through sequence weighting, position specific
gap penalties and weight matrix choice.
Nucleic Acids Research, submitted, June 1994.

Full Multiple alignment
 Calculate NJ Tree
 FAST algorithm for guide tree
 Bootstrap NJ Tree Number of bootstraps: 1000

Gap penalties: Blank=default

Pairwise alignments		Multiple alignment	
Gap open	<input type="text"/>	Gap open	<input type="text"/>
Gap extend	<input type="text"/>	Gap extend	<input type="text"/>

Other Parameters:

Note: enter additional parameters as a single line.

Output Clustal format with Clustal consensus sequence generation

Additional Parameters for ClustalW:

```
****General settings:****  
/QUICKTREE :use FAST algorithm for the alignment guide tree  
/NEWTREE= :file for new guide tree  
/USETREE= :file for old guide tree  
/NEGATIVE :protein alignment with negative values in matrix
```

Run ClustalW View ClustalW Doc Cancel

C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\edited\s2_e_F.as

2 total sequences

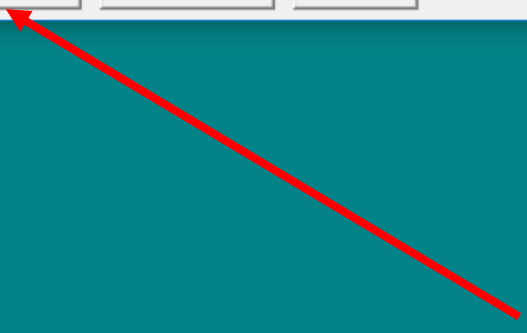
Mode: Select / Slide Selection: 0 Position: Sequence Mask: None Numbering Mask: None Start ruler at:

Font: Courier New Size: 11 Bold

AGCT GAT GAT
GGC CAT CAT

Scroll speed slow

	10	20	30	40	50	60
s2_e-Primer1	AACCGCTCTCAGAA	TATTAATTCGAAC	CAGAGTTAGGTC	CAACCTGGGAG	TTTATTGGAAAT	GATCAAAI
s2_e-Primer2	TTATATTTTATCT	TTCGGTGCTTG	ATCTGGGAGGG	TGGGAACCGCT	CCTCAGAA	TATTAATTCGAAC



Click on 'Run ClustalW'

- Minimize Alignment
- Minimize alignment to mask
- Sequence Identity Matrix
- Create Consensus Sequence**
- Entropy (H(x)) plot
- Toggle Translation Ctrl+T
- Positional Nucleotide Numerical Summary
- Positional Amino Acid Numerical Summary
- Positional frequency summary (any characters, ~- are gaps)
- Find Conserved Regions
- Similarity Matrix (for alignment and shading) >
- Strip columns containing gaps
- Plot identities to first sequence with a dot
- Information scan
- Plot a bitmap representing the alignment (1 pixel per base)
- Create IUPAC consensus in selected cums (nucleotide only, no threshold)
- Flush the alignment (pad ends with gaps to longest sequence)
- Align pipes throughout alignment
- Align spaces throughout alignment

In the new window, select both sequences again;
Create consensus sequence

The screenshot displays two windows of the BioEdit Sequence Alignment Editor. The top window, titled "C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\edited\s2_e_F.as", shows two aligned DNA sequences: "s2_e-Primer1" and "s2_e-Primer2". The bottom window, titled "Untitled", shows the same two sequences with a consensus sequence being generated, indicated by a dashed line and the sequence "AACCGCTCTCAGAAATATTAATTCGAACAGAGTTAGGTC AACCTGGGA".

BioEdit Sequence Alignment Editor

File Edit Sequence Alignment View Accessory Application RNA World Wide Web Options Window Help

- New Alignment Ctrl+N
- Open ... Ctrl+O
- New from Clipboard
- RNAMotif >
- New Text
- Open As Text
- Save Ctrl+S
- Save As...
- Retrieve sequences from GenBank or GenPept
- Copy file name to clipboard
- Export >
- Import >
- Import from Clipboard
- Merge into Alignment based on a Reference Sequence
- Merge from Clipboard
- Align and merge into alignment using top sequence as an alignment reference
- Append Alignment
- Go to GenBank
- Close
- Batch ABI to SCF trace file conversion
- Batch Export of Raw Sequence Trace Data
- Revert to Saved Ctrl+R
- Graphic View
- Print Window
- Print Alignment as Text Ctrl+P
- Untitled
- C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\edited\s2_e_R.fas
- C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\edited\s2_e_F.fas
- Untitled1
- Exit

C:\Users\alina\Dropbox\alina\UMD\Lab\Isopods_DNA barcoding\Nina\Seq_results_020119\edited\s2_e_F.fas

2 total sequences

Mode: Select / Slide Selection: 0 Position: Sequence Mask: None Numbering Mask: None Start ruler at: 1

I D I D G-D +

s2 e-Primer1 AACCGCTCTCAGAAATATTAATTCGAACAGAGTTAGGTC AACCTGGGAGTTTATTGGAAATGATCAAAI

s2 e-Primer2 TTATATTTTATCTTCGGTGCTTGATCTGGGAGGGTGGGAACCGCTCTCAGAAATATTAATTCGAACAGAG

Untitled

3 total sequences

Mode: Select / Slide Selection: 0 Position: Sequence Mask: None Numbering Mask: None Start ruler at: 1

I D I D G-D +

s2 e-Primer1 -----AACCGCTCTCAGAAATATTAATTCGAACAGAGTTAGG

s2 e-Primer2 TTATATTTTATCTTCGGTGCTTGATCTGGGAGGGTGGGAACCGCTCTCAGAAATATTAATTCGAACAGAGTTAGG

Consensus TTATATTTTATCTTCGGTGCTTGATCTGGGAGGGTGGGAACCGCTCTCAGAAATATTAATTCGAACAGAGTTAGG

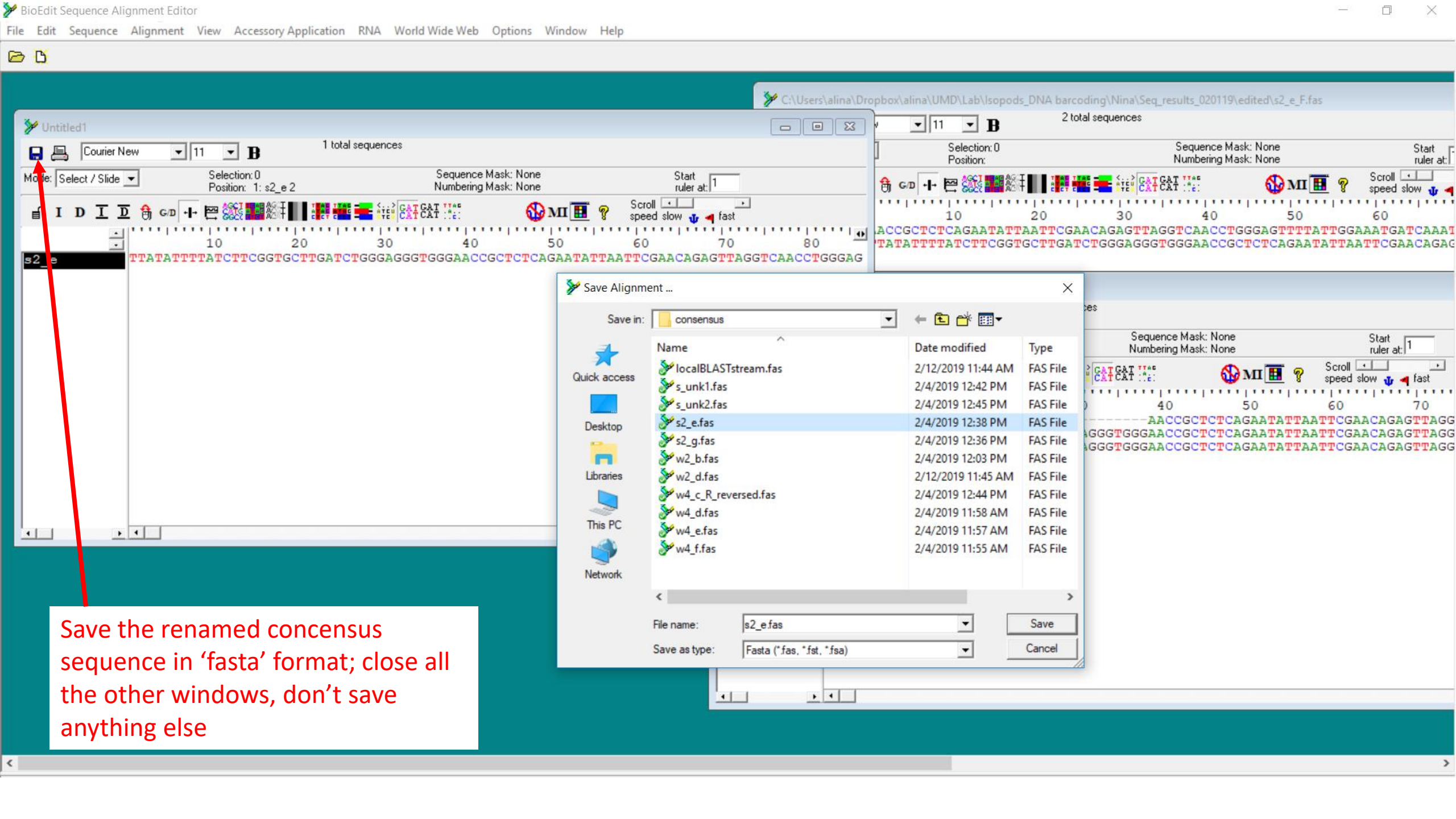
Select consensus sequence and open a new alignment window

Copy the consensus sequence in the new alignment window

The screenshot shows the main BioEdit window with a single sequence named 'Untitled1'. The consensus sequence is displayed as: `TTATATTTTATCTTCGGTGCCTTGATCTGGGAGGGTGGGAACCGCTCTCAGAAATTAATTCGAACAGAGTTAGGTC AACCTGGGAG`. An 'Edit Consensus' dialog box is open, showing the same sequence with a name field containing 's2_e'. The dialog includes fields for Name, Length (648), True Length (648), Font Size (0), and Sequence (1 out of 1). It also has checkboxes for 'Lock sequence' and 'Overwrite', and buttons for 'Apply', 'Apply and Close', and 'Cancel'. A red arrow points from the 'Consensus' label in the main window to the 'Apply and Close' button in the dialog box.

This screenshot shows multiple alignment windows in BioEdit. The top window, titled 'Untitled1', shows 11 sequences. Below it, another window shows 2 total sequences, and a third window shows 3 total sequences. Each window displays sequence alignments with a ruler at the bottom. The sequences are color-coded by base (A, C, G, T). A red arrow points from the 'Apply and Close' button in the dialog box to the 'Consensus' label in the main window.

Then double click on the consensus sequence; rename it in the new window; click on 'Apply and Close'



Save the renamed consensus sequence in 'fasta' format; close all the other windows, don't save anything else